S. P. Mandali's Ramnarain Ruia Autonomous College

(Affiliated to University of Mumbai)



Syllabus for

Program: MSc II

Program Code: RPSBCH

(Credit Based Semester and Grading System for academic year 2021–2022)



PROGRAM OUTCOMES

РО	PO Description		
	A student completing Master's Degree in SCIENCE program will be able to:		
PO 1	Demonstrate in depth understanding in the relevant science discipline. Recall, explain, extrapolate, and organize conceptual scientific knowledge for execution and application and also to evaluate its relevance.		
PO 2	Critically evaluate, analyse, and comprehend a scientific problem. Think creatively, experiment and generate a solution independently, check and validate it and modify if necessary.		
PO 3	Access, evaluate, understand, and compare digital information from various sources and apply it for scientific knowledge acquisition as well as scientific data analysis and presentation.		
PO 4	Articulate scientific ideas, put forth a hypothesis, design and execute testing tools and draw relevant inferences. Communicate the research work in appropriate scientific language.		
PO 5	Demonstrate initiative, competence, and tenacity at the workplace. Successfully plan and execute tasks independently as well as with team members. Effectively communicate and present complex information accurately and appropriately to different groups.		
PO 6	Use an objective, unbiased and non-manipulative approach in collection and interpretation of scientific data and avoid plagiarism and violation of Intellectual Property Rights. Appreciate and be sensitive to environmental and sustainability issues and understand its scientific significance and global relevance.		
PO 7	Translate academic research into innovation and creatively design scientific solutions to problems. Exemplify project plans, use management skills, and lead a team for planning and execution of a task.		
PO 8	Understand cross disciplinary relevance of scientific developments and relearn and reskill so as to adapt to technological advancements.		



PROGRAM SPECIFIC OUTCOMES

PSO	Description				
	A student completing Master's Degree in SCIENCE program in the subject of BIOCHEMISTRY will be able to:				
PSO 1	Acquire necessary knowledge and skills to undertake a career in research, either in industry or in an academic set up.				
PSO 2	Compare and contrast the breadth and depth of scientific knowledge in the broad range of fields including Protein biochemistry, Bioenergetics, Diagnostic Biochemistry, Hormonal Biochemistry, Molecular Biology, Nutritional Biochemistry, and Nanotechnology.				
PSO 3	Extrapolate and comprehend the regulatory role of metabolic processes and understand the underlying cause of metabolic disorders				
PSO 4	Acquire thorough knowledge of Biochemical Techniques, Advanced Immunology, Physiology, Genetic Engineering, and Biotechnology				
PSO 5	Describe and express the biochemical basis of human diseases, protein structure and conformation, non-invasive diagnostics, clinical research, and its importance in drug development. Usage of this knowledge further for multitude of laboratory applications.				
PSO 6	Integrate and apply the techniques in Biophysics, Analytical Biochemistry, Clinical biochemistry, Microbiology, Molecular Biology and Basics in Bioinformatics				
PSO 7	Gain proficiency in laboratory techniques in both Biochemistry and Molecular Biology, and be able to apply the scientific method to the processes of experimentation and Hypothesis testing				
PSO 8	Develop and enhance skills & improve employability through academic, research and internship opportunities				
PSO 9	Gain exposure to basic research through the provision of PG research based project.				
PSO 10	Learn to work as a team as well as independently to compile and interpret Biological data, carry out Research investigations and draw conclusions				



PROGRAM OUTLINE

YEAR	SEM	COURSE	COURSE TITLE	CREDITS	
		RPSBCH301	Biochemistry of Metabolism	4	
		RPSBCH302	Medical biochemistry	4	
		RPSBCH303	Molecular biology	4	
		RPSBCH304	Biology of diseases & Clinical	4	
	Ш		Research	V .	
		RPSBCHP301	Practicals based on RPSBCH301	2	
		RPSBCHP302	Practicals based on RPSBCH302	2	
		RPSBCHP303	Practicals based on RPSBCH303	2	
		RPSBCHP304	Practicals based on RPSBCH304	2	
MSc II		RPSBCH401	Human Physiology & Developmental	4	
			Biology		
		RPSBCH402	Endocrinology	4	
		RPSBCH403	Genetic Engineering & Biotechnology	4	
	IV	RPSBCH404	Advanced Immunology	4	
		RPSBCHP401	Discoul Co.	4	
		RPSBCHP402	- Dissertation		
		RPSBCHP403	Practicals based on RPSBCH403	2	
		RPSBCHP404	Practicals based on RPSBCH404	2	
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Semester III

Course Code: RPSBCH301

Course Title: Biochemistry of Metabolism

Academic year 2021-22

COURSE OUTCOMES:

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COURSE OUTCOME	DESCRIPTION
CO 1	Understand the major catabolic and anabolic pathways in
CO 1	metabolism of carbohydrates, lipids, amino acids and nucleotides.
CO 2	Describe regulatory mechanisms that control the metabolic
CO 2	pathways.
CO 3	Realize the Influence of Diet and hormonal signalling on metabolic
003	pathways.
CO 4	Learn Biochemical functions and integrated metabolism of in brain,
004	digestive system, liver, red cell, muscle and adipocyte.
CO 5	Illustrate the molecular mechanisms underlying major inherited
003	diseases of metabolism.
	Understand the relationship between the properties of
CO 6	macromolecules and cellular activities, cell metabolism and chemical
	composition.



Course	Unit	Course/ Unit Title	Credits/
Code/		Biochemistry of Metabolism	Lectures
Unit		RPSBCH301	4 Credits
	1	Carbohydrate Metabolism	15L
	1.1	Schematic representation of Glycolysis & Kreb's	2L
		cycle	
	1.2	Gluconeogenesis: Pathway and its Regulation	1L
	1.3	Glycogen Metabolism: Synthesis, breakdown,	3L
		mechanisms of control of glycogen metabolism	
		Direct Allosteric Control of Glycogen	
		Phosphorylase and Glycogen Synthase, Covalent	
		Modification of Enzymes by Cyclic Cascades,	
		Hormonal regulation, Maintenance of Blood	
		Glucose Levels	
ı	1.4	Futile cycle, Rapoport Luebering cycle, Cori cycle, Glucose-Alanine cycle & their significance	2L
	1.5	Shuttles-Malate-Aspartate shuttle & Glycerol	3L
		phosphate shuttle	_
	1.6	Uronic acid pathway (biosynthesis, degradation &	
		its significance),	
		Galactose and fructose metabolism; Sorbitol	
		pathway	
	1.7	Biosynthesis of oligosaccharides and glycoproteins	1L
	1.8	Synthesis of carbohydrates in plants	3L
	1.8.1	Calvin cycle and its regulation, regulated synthesis	
		of starch and sucrose, photorespiration, C4 and	
	1/1	CAM pathways, Glyoxylate pathway, synthesis of	
		cell wall polysaccharides	
	2	Amino acid metabolism	15L
	2.1	Reactions of amino acids: Deamination,	3L
O hy		Transamination, Decarboxylation,	
		Transmethylation, Transdeamination,	
	2.2	Ammonia formation, transport and detoxification in	
l II		brain and liver	
	2.3	Kreb's bicycle, Urea cycle & its regulation.	2L
		Inherited defects of urea cycle	
	2.4	Glucogenic and ketogenic amino acids.	
	2.5	Metabolism of one carbon units. Disorders of	3L
		amino acids metabolism, phenylketonuria,	
		alkaptonuria, maple syrup urine disease,	



		methylmalonic acidemia (MMA), homocystinuria	
		and Hartnup's disease	
	2.6	Biosynthesis of amino acids	2L
		Overview of amino acid synthesis. Biosynthesis of	
		non-essential amino acids and its regulation	
	2.7	Precursor functions of amino acids	1L
	2.7.1	Biosynthesis of creatine and creatinine, polyamines	4L
		(putresine, spermine, spermidine), catecholamines	
		(dopamine, epinephrine, norepinephrine) and	
		neurotransmitters (serotonin, GABA).	$\mathcal{C}_{\mathcal{X}}$
	3	Lipid metabolism	15L
	3.1	Fatty acid oxidation	4L
	3.1.1	Fatty acid transport to mitochondria, β-oxidation of	
		saturated, unsaturated, odd and even numbered	
		and branched chain fatty acids, regulation of fatty	
		acid oxidation, peroxisomal β-oxidation, ω	
		oxidation, ketone bodies metabolism, ketoacidosis	
	3.2	Fatty acid synthesis	2L
	3.2.1	Transport of mitochondrial Acetyl Co A to cytosol,	
		Fatty acid synthase complex, Synthesis of	
III		saturated, unsaturated, odd and even chain fatty	
		acids and regulation.	
	3.2.2	Biosynthesis of eicosanoids, cholesterol, steroids	2L
		and isoprenoids	
	3.2.3	Synthesis of prostagladins, leukotrienes and	3L
		thromboxanes. Synthesis of cholesterol, regulation	
		of cholesterol synthesis. Synthesis of steroids and	
		isoprenoids.	
	3.2.3	Biosynthesis of glycerophospholipids and	2L
	N	sphingolipids	
	3.4	Lipid storage diseases	2L
	4	Nucleic Acid Metabolism & Integration of	15L
1/3		Metabolism	
O.K.	4.1	Biosynthesis of purine and pyrimidine nucleotides	3L
	4.1.1	De novo synthesis of purine and pyrimidine	
		nucleotides, regulation and salvage pathways.	
IV		Deoxyribonucleotides and synthesis of nucleotide	
	4.4.0	triphosphate	01
	4.1.2	Biosynthesis of deoxyribonucleotides and its	2L
	4.0	regulation, conversion to triphosphates,	41
	4.2	Biosynthesis of coenzyme nucleotides	1L
	4.3	Degradation of purine and pyrimidine nucleotides	2L



4.4	Digestion of nucleic acids, degradation of purine	1L
	and pyrimidine nucleotides. Inhibitors of nucleotide metabolism.	
4.5	Disorders of purine and pyrimidine metabolism –	3L
	Lesch-Nyhan, syndrome, Gout, SCID, adenosine	
	deaminase deficiency	
4.6	Integration of metabolism	1L
4.6.1	Integration of metabolic pathways (carbohydrate,	2L
	lipid and amino acid metabolic pathways), tissue	
	specific metabolism (brain, muscle, and liver).	CX
	Practicals – RPSBCHP301	2 Credits
1	Estimation of glucose by the GOD-POD method	
2	Study of glucose metabolism using handheld	
	glucometer	
3	Estimation of amino acids by ninhydrin	
4	Estimation of serum urea by diacetyl monoxime	
	method	
	Estimation of serum creatinine by alkaline picrate	
5	method	
_	Assay of glutamate dehydrogenase	
6	Estimation of serum uric acid by phosphotungstic	
7	acid method (Caraways method)	
_	Use of softwares to understand metabolism –	
8	KEGG, Ecocyc, Metacyc, Biocyc	

- Lehninger: Principles of Biochemistry (2013) 6th ed., Nelson, D.L. and Cox, M.M., W.H. Freeman and Company (New York), ISBN:13:978-1-4641-0962-1 / ISBN:10:1-4641-0962-1.
- 2. Textbook of Biochemistry with Clinical Correlations (2011) 7th ed., Devlin, T.M., John Wiley & Sons, Inc. (New Jersey), ISBN:978-0-470-28173-4.
- 3. Biochemistry (2012) 7th ed., Berg, J.M., Tymoczko, J.L. and Stryer L., W.H. Freemanand Company (New York), ISBN:10:1-4292-2936-5, ISBN:13:978-1-4292-2936-4.
- 4. Principles of Biochemistry by G. Zubay, W. Parson, D.
- 5. Biochemistry Voet, D. and Voet, J.G. John Wiley & Sons, Inc. USA.



Course Code: RPSBCH302

Course Title: Medical Biochemistry

Academic year 2021-22

COURSE OUTCOMES:

COURSE	DESCRIPTION
CO 1	The student will learn and understand the basics of circulatory
	system including haematopoiesis, homeostasis, and diseases of
	blood.
CO 2	Explain the fundamentals, composition and significance of Body
	fluids
CO 3	To get acquainted with the role of enzymes in diagnosis of various
	diseases.
CO 4	The student will be aware of the organ function tests available for
	detection of their functionality
CO 5	Illustrate Pathophysiology of certain disorders related to certain
	organs & Clinical tests available for detection
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Course	Unit	Course/ Unit Title	Credits/
Code/		Medical Biochemistry	Lectures
Unit		RPSBCH302	4 Credits
	1	Haematopoiesis & related disorders	15L
	1.1	Introduction to Haematopoiesis	1L
	1.1.1	Erythropoiesis - Stages of development of	2L
		erythrocytes, Precursors of RBCs, Factors	
		influencing erythropoiesis	
	1.1.2	Role of erythropoietin	
	1.2	Haemoglobin (Hb)-Features, varieties, combination	4L
		of Hb with gases, Haeme-haeme interactions	
	1.2.1	Biosynthesis of Haemoglobin (with structures)	
	1.2.3	Biochemical pathway for Porphyrin synthesis,	
		formation of Haeme	
I	1.3	Haeme catabolism	2L
	1.4	Haemoglobinopathies	2L
	1.4.1	Genetics basis of haemoglobinopathies - Sickle	
		cell anemia, Thalassemia – alpha (Subtypes of	
		alpha thalassemia) & beta	
	1.5	Blood gas analysis	1L
	1.5.1	Leucopoiesis, Leucocytosis and factors	2L
		responsible, Leukopenia, Thrombopoiesis,	
		Thrombocytopenia	
	1.6	Molecular mechanism of blood coagulation, role of	1L
		vitamin K in coagulation, anticoagulant and	
	-	fibrinolytic systems, haemophilia and thrombosis	
	2	Biochemistry of body Fluids & related	15L
		disroders	
	2.1	Plasma	3L
	2.1.1	Composition, Separation of plasma proteins –	
		Salting out, Cohn's fractionation, Electrophoresis	
U	2.1.3	Characteristics, functions and clinical significance	
		of plasma proteins	
	2.2	Proteinuria – types & causes	1L
	2.3	Bile	4L
	2.3.1	Bile pigments- Biliverdin and Bilirubin	
	2.3.2	(formation, transport, conjugation in liver cells,	
		secretion in bile, excretion)	



		1	
	2.3.3	Clinical importance of bile formation (concept -	
		index of rate of haeme catabolism, shunt	
		hyperbilirubinaemia, bilirubin encephalopathy)	
	2.4	Lymph- Formation, composition and circulation	1L
	2.5	CSF- Composition, Appearance, Pressure of CSF	2L
		and Biochemical Changes in CSF and its clinical	
		significance	
	2.6	Synovial fluid- Composition, Classification and	1L
		Clinical significance – synovial fluid viscosity,	
		pathology	CX_{\bullet}
	2.6.1	Analysis – mucin clot test, microscopic analysis	JL.
	2.7	Pericardial fluid- Composition, Function &	2L
		Pericardiocentesis, Pericardial effusion, Ischemic	
		heart disease	
	3	Enzymes & isoenzymes of clinical importance	15L
	3.1	Introduction, Possible mechanisms responsible for	4L
		abnormal enzyme levels	
	3.2	Value of serum enzyme assay in clinical practice	
	0.2	Enzyme assays of clinical significance	
	3.3	Myocardial infarction – Creatine phosphokinase, S-	4L
	0.0	GOT, LDH. Important cardiac markers useful in	76
III		diagnosis of acute myocardial infarctions	
	3.4	GI tract diseases - Serum amylase, serum lipase	2L
	3.5	Liver diseases – Serum transaminases	ZL
	3.6		2L
		Muscle diseases – Serum aldolase, serum CPK	ZL
	3.7	Bone diseases – Serum alkaline phosphatases	41
	3.8	Malignancies – Acid phosphatase, β-glucuronidase	1L
	3.9	Value & clinical significance of isoenzymes of –	2L
	2	LDH, CPK, Alkaline phosphatase	4=1
	4	Organ Function Tests	15L
	4.1	Renal Function test Preliminary investigations	2L
		Classification of renal function tests – Tests based	
		on glomerular filtration, Tests to measure Renal	
M.		Plasma Flow, Tests based on tubular function	
IV	4.2	Liver Function test	2L
]		Tests based on abnormalities of bile pigment	
		metabolism - VD Bergh Reaction, Determination of	
		Serum Bilirubin	
		Tests based on liver's part in carbohydrate	
		metabolism – Glucose tolerance test, Galactose	
		tolerance test, Fructose tolerance test	



	4.3	Tests based on changes in plasma proteins –	3L
		Determination of Total Plasma Proteins and	
		Albumin and Globulin and A:G Ratio, Estimation of	
		Plasma Fibrinogen, Flocculation Tests, Amino	
		acids in urine	
	4.4	Tests based on abnormalities of lipids -	2L
		Determination of serum cholesterol and ester	
		cholesterol and their ratio, Determination of faecal	
		fats	
	4.5	Tests based on detoxicating function of liver -	2L
		Hippuric acid synthesis test	
	4.6	Gastric Function test – Fractional gastric analysis,	2L
		stimulation tests	
	4.7	Thyroid Function test	2L
		Tests based on primary function of thyroid, Tests	
		measuring blood levels of thyroid hormones	
		Practicals – RPSBCHP302	2 Credits
	1	Enumeration of Blood cells: RBC and WBC	
		counting	
	2	Qualitative analysis of bile	
	3	Estimation of serum electrolytes	
	4	Estimation of blood glucose by ortho-toluidine	
		mono step method	
	5	Estimation of serum phosphorus by Fiske	
		Subbarow method	
	6	Estimation of serum Calcium by Trinder's method	
	7	Estimation of serum iron by dipyridyl method	
	8	Clinical analysis of CSF – glucose, proteins,	
		chlorides Liver Function Tests –	
	9	a) Estimation of serum ALT and AST	
		b) Estimation of total and direct bilirubin	
	12.	c) Estimation of serum alkaline phosphatase	
6/3		d) Estimation of total proteins, albumin and	
Oh,		determination of A/G ratio	
		e) Estimation of serum albumin by Bromocresol	
		Green (BCG) binding method	
	10	Renal Function tests	
		a) Creatinine clearance test	
		b) Urea clearance test	
	11	Pancreatic Function Test	
	''	a) Estimation of serum amylase activity	
		b) Glucose Tolerance Test	
		S) Sidosoc Folciarios Fost	



12	2	Estimation of serum total cholesterol and HDL	
		cholesterol	
13	3	Estimation of triglycerides	

- 1. Textbook of Biochemistry with Clinical Correlations (2011) Devlin, T.M. John Wiley & Sons, Inc. (New York), ISBN: 978-0-4710-28173-4.
- 2. Introduction to Human Physiology (2013) 8th edition; Lauralee Sherwood. Brooks/Cole, Cengage Learning.
- 3. The World of the cell, 7th edition (2009)
- 4. Genetics (2012) Snustad and Simmons
- 5. Urinalysis and Body Fluids by Susan King Strasinger & Marjorie Schaub Di Lorenzo, 6th Edition
- 6. Graff's Textbook of Urinalysis and Body Fluids Lillian A. Mundt & Kristy Shanahan, 2nd Edition
- 7. Fundamentals of the study of urine and body fluids Nancy A. Brunzel, 3rd Edition, Elsevier
- 8. A Textbook of Medical Biochemistry MN Chatterjea & Rana Shinde, 8th Edition, Jaypee Publication
- 9. Clinical Biochemistry Metabolic and Clinical Aspects by William J. Marshall, Márta Lapsley, Andrew Day, Ruth Ayling
- 10. Cooper, G.M. and Hausman, R.E. 2009 The Cell: A Molecular Approach. 5th edition. ASM Press & Sunderland, Washington, D.C.; Sinauer Associates, MA Principles of Biochemistry by G. Zubay, W. Parson, D.



Course Code: RPSBCH303

Course Title: Molecular biology

Academic year 2021-22

COURSE OUTCOMES:

COURSE	DESCRIPTION
OUTCOME	
CO 1	Understand the mechanism of cell cycle, relationship of cell cycle
	and programmed cell death via intracellular and extracellular control
	mechanisms
CO 2	The students will learn about nucleic acid as genetic information
	carriers, Possible modes of replication, and roles of helicase,
	primase, gyrase, topoisomerase, DNA Polymerase, DNA ligase,
	and Regulation of replication.
CO 3	The student will be able to illustrate the mechanism of prokaryotic
	and eukaryotic replications
CO 4	The student will learn & understand different types of mutations,
	agents causing mutations and disorders resulting from mutations.
CO 5	Comprehend the mechanism and regulation of transcription in
	prokaryotes along with Reverse transcription.
CO 6	The student will be able to describe synthesis of protein from gene
	with the help of regulatory protein
CO 7	The student will be able to explain the Post transcriptional &
	translational modifications & their significance in stability



Course	Unit	Course/ Unit Title Credits/			
Code/		Molecular Biology	Lectures		
Unit		RPSBCH303	4 Credits		
	1	Prokaryotic and eukaryotic Transcription &	15L		
		Post-transcriptional Modifications			
	1.1	Overview of Transcription	1L		
	1.2.1	Role of Prokaryotic RNA polymerase and	4L		
		promoter; Upstream regulatory sequences			
	1.2.2	Stages of transcription: Initiation, elongation and			
	termination (Rho dependent & Rho independent)				
	1.2.3	Significance of Sigma factor, Concept of Abortive			
		initiation			
	1.3	Transcription in eukaryotes	3L		
	1.3.1	Role of promoter & regulatory elements			
	1.3.2	Eukaryotic RNA polymerases			
	1.3.3	Mechanism of RNA transcription in eukaryotes -			
-		Formation of pre-initiation complex, initiation,			
	elongation and termination, Phosphorylation of				
		RNAPII	1L		
	1.4 RNA Modification				
	1.4.1	Mechanism of addition of 5'-cap & formation of poly			
		A tail, 5'-cap of snRNA	1L		
	1.4.2 Molecular mechanism of mRNA Splicing				
	1.4.3 RNA Processing of rRNA & tRNA1.4.4 RNA editing - Base modifications, role of snoRNA		1L		
			1L		
	1.5	Role of Inhibitor -Rifampicin, Actinomycin D,	1L		
		α-Amanitin			
	1.6	Reverse transcription (Mechanism, significance &	2L		
	2	application)	451		
O'Ay,	2	Prokaryotic and eukaryotic Translation & Post-	15L		
	2.1	translational Modifications Canadia basis of protein bissynthesis. Canada of	41		
	2.1	Genetic basis of protein biosynthesis – Concept of	4L		
l II		structural gene & Protein, Characteristics of Genetic code			
"	2.2	Ribosome assembly & structure (Comparison			
	2.2	between prokaryotic & eukaryotic ribosome)			
	2.3	tRNA – structural features and tRNA synthetase,	2L		
	2.3	initiator tRNA, activation of amino acids	ZL		
		minator titiva, activation of amino acius			



	Mechanism of translation in prokaryotes: Initiation,	3L	
		elongation & termination	
	2.4.2	Concept of Polyribosome	
	2.5	Mechanism of eukaryotic translation: Initiation,	2L
		elongation & termination	
	2.6	Inhibitors of translation (prokaryotes & eukaryotes)	1L
	2.7	Post translational modifications of proteins	1L
	2.8.1	Signal hypothesis	2L
	2.8.2	Role of signal peptide & its role in Protein sorting	<.
	2.9	Protein localization in Nucleus	. (.)
	3	Gene regulation in prokaryotes	15L
	3.1.1	Principles of gene regulation, Constitutive &	3L
		inducible genes, one cistron-one subunit concept	
	3.1.2	Negative and positive regulation	
	3.2	Concept of operons, regulatory proteins, activators,	2L
		repressors, DNA binding domains, allosteric site	
III	3.3	Lac operon – Structure, inducers (allolactose,	3L
		IPTG), Negative control & Positive control of lac	
		operon	
	3.4	Tryptophan operon – structure & regulation	2L
	3.5	Arabinose operon – structure, function & regulation	2L
	3.6	Regulatory RNAs in bacteria, small RNA and	3L
		riboswitches	
	4	Gene regulation in eukaryotes	15L
	4.1.1	Role of regulatory transcription factors in	2L
		eukaryotic gene regulation-general TF and	
		Regulatory TF, TFIID and Mediator	
	4.1.2	Modulation of the function of regulatory	2L
		transcription factors	
	111	Ligand modification	
. 5		Protein protein interaction	
	//	Covalent modification	
IV	4.2.1	Gene regulation by chromatin remodelling –	4L
O'A.		removal of histone octamer nucleosome structure,	
		shifting of the nucleosome and changing the	
		canonical subunits of histone octamer for the non-	
		canonical subunits	
	4.2.2	Histone acetylation and deacetylation, Gene	
		silencing – DNA methylation	
	4.2.3	Regulation of galactose metabolism in yeast	2L
	4.2.4	Regulation by phosphorylation of nuclear	2L
		transcription factors	



4.3	Regulatory RNAs in eukaryotes: synthesis and	3L
	mechanism of siRNA and miRNA	
	Practicals – RPSBCHP303	2 Credits
1	To hydrolyze DNA and separate nucleotide bases	
	by paper chromatography	
2	Isolation of chromosomal DNA from E coli cells	
3	Qualitative Analysis of DNA by AGE	
4	Designing of Oligonucleotide primers for PCR	
5	Amplification of a DNA fragment by PCR	
6	Cytochemical staining of RNA by Methyl Green	CX
	Pyronin	
7	Total RNA isolation from Bacterial Cells	

- Molecular Biology of the Gene (2008) 6th ed., Watson, J.D., Baker, T.A., Bell, S.P., Gann, A., Levine, M. and Losick, R., Cold Spring Harbor Laboratory Press, Cold Spring Harbor (New York), ISBN:0-321-50781 / ISBN: 978-0-321-50781-5.
- 2. Cooper, G.M. and Hausman, R.E. 2009 The Cell: A Molecular Approach. 5th edition. ASM Press & Sunderland, Washington, D.C.; Sinauer Associates, MA.
- 3. Karp, G. 2010 Cell and Molecular Biology: Concepts and Experiments. 6 edition. John Wiley & Sons. Inc.
- 4. Alberts, B., Johnson, A., Lewis, J., and Enlarge, M. 2008 Molecular Biology of the Cell.
- 5. Genetics: Analysis & principles by Robert brooker 5th edition, McGraw-Hill Education, 201
- 6. Molecular Cell Biology by Lodish, Berk, Matsudaira, Kaiser, Krieger, Scott, Zipursky, Darnell



Course Code: RPSBCH304

Course Title: Biology of diseases & Clinical Research

Academic year 2021-22

COURSE OUTCOMES:

COURSE	DESCRIPTION				
OUTCOME					
CO 1	The student will able to understand the cooperative relationship				
	between Biochemistry and Medicine				
CO 2	Learn the factors in the spread of infectious diseases & Explain				
	biology and pathogenesis of infectious agents				
CO 3	Describe the Immune responses of body against various				
	pathogenic organisms				
CO 4	Recognize the Biochemical aspect of cancer, Assays for diagnosis				
	& treatment				
CO 5	Understand the Physiology of cardiovascular diseases				
CO 6	Realize the importance of drug development through clinical				
	research				
Q.A.M.					



Course	Unit	Course/ Unit Title	Credits/
Code/		Biology of diseases & Clinical Research	Lectures
Unit		RPSBCH304	4 Credits
	1	Infectious diseases	15L
	1.1	Bacterial infections: Tetanus, Diphtheria,	3L
		Tuberculosis, Typhoid, Cholera	
	1.2	Viral infection: Measles, Mumps, influenza, HIV	
	1.3	Protozoan: Malaria and Trypanosomiasis	3L
	1.4	Parasitic infection: Leishmaniasis	
	1.5	Treatment of infectious agents	
	1.5.1	Characteristics of an ideal chemotherapeutic agent	2L
I	1.6	Mode of action of antibiotics on	
	1.6.1	Cell wall (Penicillin and Cephalosporins)	1L
	1.6.2	Cell Membrane (Polymyxin and Imidazole)	1L
	1.6.3	Protein Synthesis (Streptomycin, Tetracycline and	1L
		Chloramphenicol)	
	1.6.4	Nucleic acid (Quinolones, Nalidixic acid, Rifamycin)	1L
	1.6.5	Enzyme inhibitors (Trimethoprim)	1L
	1.7	Mechanisms of drug resistance- evolution,	2L
	1.7	pathways and origin	
	2	Cancer	15L
	2.1	Genetic basis of cancer	1L
	2.2	Experimental evidenced for transformation of cell	
	2.3	Oncogenes	3L
	2.3.1	Identification of chromosomal oncogene	
2	2.3.2	Gain of function mutation	
	2.3.3	Conversion of proto-oncogene to oncogene	2L
1/1/4	2.3.4	Missense mutation, Gene amplification,	
H		chromosomal translocation, viral integration	
	2.5	Tumor suppressor gene- Role of p53 and RB gene	2L
	2.6	Assays – Trypan blue exclusion method, MTT	3L
		assay, Soft Agar Colony Formation Assay	
	2.7	Molecular profiling for classification of cancer,	
	2.8	DNA microarray	
	2.9	Cancer therapy- Antimetabolites, Chemotherapy	4L
		(purine & pyrimidine analog), Demethylating agents	
	2.9.1	Cancer immunotherapy	



	3	Cardiovascular diseases	15L	
	3.1	Definition; The origin of cardiovascular diseases	2L	
		(electrical, structural and circulatory) and types of		
		CVDs		
	3.2	Defining the broad spectrum of Ailments	3L	
	3.3	Stages of CVDs		
III	3.4	Molecular basis of CVDs - hypertension, coronary	5L	
•••		heart (artery) disease, cerebrovascular disease,		
		cardiomyopathy, cardiac hypertrophy,	<.	
		atherosclerosis, myocardial infarction.		
	3.5	Diagnosis and Treatment strategies: screening	5L	
		methods; Current treatment modalities and their		
		advantages and disadvantages, major side effects,		
		Challenges of treatment. biomarkers for CVDs		
	4	Clinical research	15L	
	4.1	Introduction of Clinical Research	1L	
	4.1.1	Clinical Trial Phases & Evaluation	2L	
		Role of Placebo		
		Interpretation of clinical data, Meta-analysis		
	4.1.2	Pharmacological Principal of Clinical Research	2L	
		Drug Development And Launch		
	4.1.3	Clinical Trial Design and Project Managements	1L	
	4.2	Drug Invention; New Drug Development and Drug	2L	
		Assay		
	4.2.1	Animal Toxicity studies: Systemic toxicity studies,		
		Local toxicity studies & Specialised toxicity studies		
IV		Interpretation of animal data, Subjective responses		
	4.2.2	Drug Assay:	2L	
		1. Chemical		
	11	2. Biological – Indication, Principle, Types &		
7.0		Biostandardisation		
	4.0	Immunological	41	
	4.3	Radio-receptor assays & ELISA	1L	
		Essential documents in Clinical Research and	4L	
	1 1 1	Regulatory Requirements		
	4.4.1	IND Application		
		NDA Application		
	4.4.3	Informed Consent process and Documentation Clinical Study Papert		
	4.4.4	Clinical Study Report Practicals – RPSBCHP304	2 Credits	
	1 2	WIDAL test – Qualitative & Quantitative Bioassay of penicillin/ampicillin		
	3	Determination of MIC of antibiotic		
	J	Determination of Mile of antibiotic		



	Separation of isoenzymes of LDH by	
4	electrophoresis.	
	Visualization of cells by methylene blue	
5	Study of viability of cells using Neutral red Assay	
6	Spectroscopic estimation of aspirin hydrolysate	
7	Case studies on clinical research	
8		

- Textbook of Biochemistry with Clinical Correlations (2011) Devlin, T.M. John Wiley & Sons, Inc. (New York), ISBN: 978-0-4710-28173-4.
- 2. Immunology: A Short Course (2009) 6th ed., Coico, R and Sunshine, G., John Wiley& sons, Inc (New Jersey), ISBN: 978-0-470-08158-7
- 3. Biochemistry (2012) 7th ed., Berg, J.M., Tymoczko, J.L. and Stryer, L., W.H Freeman and Company (New York), ISBN: 13:978-1-4292-7635-1.
- 4. Genetics (2012) 6th ed., Snustad, D.P. and Simmons, M.J., John Wiley & Sons. (Singapore), ISBN: 978-1-118-09242-2.
- 5. Genetics Analysis and Principles by Robert J. Brooker
- 6. Concepts of Genetics by William S. Klug, Michael R. Cummings, Charlotte A. Spencer, Michael A. Palladino, Darrell Killian
- 7. Textbook of Pharmacology F.S.K Barar
- 8. Molecular Biology of Cancer Mechanisms, Targets, and Therapeutics by Lauren Pecorino
- 9. Clinical Trials, Second Edition_ Study Design, Endpoints and Biomarkers, Drug Safety, and FDA and ICH Guidelines Tom Brody
- 10. Early Drug Development Strategies and Routes to First-in-Human Trials by Mitchell N. Cayen
- Pharmacology and Pharmacotherapeutics by R.S. Satoskar, Nirmala N. Rege, S.D. Bhandarkar – 24th Edition
- 12. Textbook of Designing Clinical Research by Stephen B. Hulley



Modality of Assessment (SEMESTER III)

Theory Examination Pattern:

- A) Internal Assessment- 40%- 40 Marks
- B) External Examination- 60%- 60 Marks
 Semester End Theory Examination: (Deviation from the usual modality)
 Owing to the pandemic situation prevailing in 2020 and continuing in 2021, the external examinations (Semester End) may be conducted online as per the instructions/circulars received from the University of Mumbai and Maharashtra State notifications from time to time. The conventional mode of external examination will commence again only after the declaration of normalcy by the Government authorities.
 - 1. Duration These examinations shall be of 02 ½ HOURS duration.
 - 2. Theory question paper pattern:

Paper Pattern:

Question	Options	Marks	Questions Based on
Q1. A	Any 1 out of 2	03	LINUT
Q1. B	Any 2 out of 3	06	UNIT I
Q2. A	Any 1 out of 2	03	UNIT II
Q2. B	Any 2 out of 3	06	UNIT II
Q3. A	Any 1 out of 2	03	UNIT III
Q3. B	Any 2 out of 3	06	UNIT III
Q4. A	Any 1 out of 2	03	UNIT IV
Q4. B	Any 2 out of 3	06	OINIT IV
	TOTAL	60	



Practical Examination Pattern:

A) Internal Examination: 40%-40 Marks

Particulars	Practical I, II, III & IV
Journal	05
Experimental tasks	15
Total	20

B) External Examination: 60%-60 Marks

Semester End Practical Examination:

Particulars	Practical I, II, III & IV
Laboratory work	25
Viva	5
Total	3.0

Overall Examination & Marks Distribution Pattern

Semester III

Course	301			3	02		Grand Total
	Internal	External	Total	Internal	External	Total	
Theory	40	60	100	40	60	100	200
Practicals	20	30	50	20	30	50	100

Course	303			3	04		Grand Total
	Internal	External	Total	Internal	External	Total	
Theory	40	60	100	40	60	100	200
Practicals	20	30	50	20	30	50	100



Semester IV

Course Code: RPSBCH401

Course Title: Human Physiology & Developmental Biology

Academic year 2021-22

COURSE OUTCOMES:

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COURCE	DESCRIPTION
COURSE	DESCRIPTION
OUTCOME	
CO 1	The student will learn and understand the basic concepts of nutrition,
	and nutritional values of foods, and Basal metabolic rate and
	measurement of energy requirements.
CO 2	Deduce the dietary requirement of carbohydrates, lipids and
	proteins and their biological significance
CO 3	The course will also aid to learn the nutritional requirement and
	significance of dietary minerals like macroelements and
	microelements
CO 5	Understand the condition of malnutrition, its prevention, and
	recommended dietary allowances.
CO 6	Students will be learning topics like Introduction to nutrition,
	Macroelements, Microelements and Nutrigenomics, Nutritional
	disorders and Antinutritional Factors



Course	Unit	Course/ Unit Title	Credits/
Code/		Human Physiology & Developmental	Lectures
Unit		Biology	4 Credits
		RPSBCH401	
	1	Musculoskeletal system and its related	15L
	-	disorders	
	1.1	Bones- Composition, formulation, Structure and	3L
		functions	
	1.2	Bone's role in calcium homeostasis, factors	X >
		affecting bone metabolism, bone remodeling	
	1.3	Diseases related to bone - Osteogenesis	3L
		imperfecta, osteoporosis, osteomalacia, Fibrous	
		dysplasia, Osteomyelitis	
	1.4	Physiology of muscle contraction in striated and	3L
		non-striated muscle	
	1.4.1	Types of muscles: ultrastructure of smooth,	
		skeletal and cardiac muscle fibers	
	1.4.2	Muscle proteins, organization of contractile protein	
		and definition of sarcomeres	
	1.4.3	Mechanism of muscle contraction, excitation of	4L
_	1.4.4	striated muscles Energetics of muscle contraction, regulation of	
ı	1.4.4	skeletal muscle contraction	
	1.4.5	Calmodulin and its regulatory role	
	1.4.5		
	1.4.6	Diseases related to muscle - muscular dystrophies,	2L
		myositis	
	2	Cardiovascular Physiology	15L
20	2.1	Pericardium – Structure, function and clinical	2L
		significance	0.1
1/1/2	2.2	Cardiac muscle tissue, difference in atrial and	2L
O'h.	2.3.1	ventricular cardiac muscle, clinical significance Action potentials in cardiac muscle – Causes,	4L
	2.5.1	Phases	46
II	2.3.2	Velocity of signal conduction, Refractory period of	
	2.0.2	cardiac muscle	
	2.5	Function of Ca2+ ions and transverse tubules in	
		excitation-contraction coupling	
	2.6.1	Physical characteristics of the circulation	4L
	2.6.2	Basic principles of circulatory function	
	2.6.3	Interrelationships of pressure, flow and resistance	
1	1		I



	2.7	Vascular distensibility	3L
	2.8	Microcirculation – Structure and capillary system	
	3	Neurophysiology and its related disorders	15L
	3.1	Nervous system and its classification	3L
	3.2	Neuron - Structure and maintenance of neurons,	
		Functional classes of neurons	
	3.3	Non-neuronal cells and nerve tissue	
	3.4	Membrane potentials: Resting Membrane	5L
		Potential, Graded potentials, Action potential	
	3.5	Transmission of nerve impulse, role of Ca+2 in	CX
Ш		release of neurotransmitter from pre-synaptic	
•••		membrane, function of receptor proteins and	
		secondary messengers on the postsynaptic neuron	
	3.6	Chemical & electrical synapse, Post-synaptic	2L
		potentials: excitatory and inhibitory	
	3.7	Characteristics of some important	3L
		neurotransmitters (Acetylcholine, Glycine, GABA,	
		Aspartate, Glutamate, Catecholamines)	
	3.8	Disorders related to defects in neurotransmission-	2L
		Parkinson's disease, stroke, Alzheimer's disease	
	4	Developmental Biology	15L
	4.1	Basic concepts of development : Potency,	2L
		commitment, specification, induction, competence,	
	4.0	determination and differentiation	01
	4.2	Morphogenetic gradients; cell fate and cell	2L
		lineages; genomic equivalence and the	
	4.3	cytoplasmic determinants Gametogenesis, fertilization (in humans & sea	3L
IV	7.5	urchin)	JL
	4.4	Early development, cell surface molecules in	3L
		sperm-egg recognition in animals	~
- 5	4.5	Embryonic cleavage	2L
	4.6	Formation of germ layers in animals	
OHI	4.7	Sexual reproduction in plants - Gametogenesis,	3L
		double fertilization in plants	



- 1. Vander's Human Physiology (2008) 11th ed., Widmaier, E.P., Raff, H. and Strang, K.T., McGraw Hill International Publications (New York), ISBN: 978-0-07-128366-3.
- 2. Harper's Biochemistry (2012) 29th ed., Murray, R.K., Granner, D.K., Mayes and P.A., Rodwell, V.W., Lange Medical Books/McGraw Hill. ISBN:978-0-07-176-576-3.
- 3. Textbook of Medical Physiology (2011) 10th ed., Guyton, A.C. and Hall, J.E., Reed Elseviers India Pvt. Ltd. (New Delhi). ISBN: 978-1-4160-4574-8.
- 4. Fundamental of Anatomy and Physiology (2009), 8th ed., Martini, F.H. and Nath, J.L., Pearson Publications (San Francisco), ISBN: 10:0-321-53910-9 / ISBN: 13: 978-0321-53910-6.



Course Code: RPSBCH402

Course Title: Endocrinology

Academic year 2021-22

COURSE OUTCOMES:

COURSE	DESCRIPTION
OUTCOME	
CO 1	Study the historical experiments that lead to the discovery of
	various hormones
CO 2	Deeply understand the communication between the nervous
	system and the endocrine system
CO 3	Learn the structure, functions and the disorders associated with the
	various hormones starting from the pituitary hormones to the
	gonadal hormones.
CO 4	Appreciate and analyze the endocrine regulation of the various
	metabolisms such as carbohydrate metabolism, Protein
	metabolism, calcium homeostasis, menstrual cycle, pregnancy and
CO 5	menopause. Apply the knowledge of hormones in assay of hormones such as
	T3, T4 and TSH and understand the strategy behind contraception.
CO 6	Present a case study on a hormonal and a metabolic disorder
RAMIN	



Course	Unit	Course/ Unit Title	Credits/
Code/		Endocrinology	Lectures
Unit		RPSBCH402	4 Credits
	1	Introduction to Endocrinology & cellular	15L
		signalling	
	1.1	Functions of hormones and their regulation	2L
	1.2	Chemical signaling - endocrine, paracrine,	
		autocrine, intracrine and neuroendocrine	
		mechanisms	X >
	1.3	Chemical classification of hormones, transport of	1L
		hormones in the circulation and their half-lives	
	1.4	Hormone receptors - extracellular and intracellular	3L
ı	1.4.1	Receptor - hormone binding, Scatchard analysis	
	1.4.2	G protein coupled receptors, G proteins, second messengers - cAMP, cGMP, IP3, DAG, Ca ²⁺	4L
	1.5	Effector systems - adenyl cyclase, guanyl cyclase, PDE, PLC	
	1.5.1	Protein kinases (PKA, PKB, PKC, PKG)	
	1.5.2	Receptor tyrosine kinases - EGF, insulin and Ras -	4L
	1.0.2	MAP kinase cascade	7⊑
	1.5.3	Non receptor tyrosine kinase-erythropoietin	
		receptor JAK - STAT pathway	
	1.6	Steroid hormone Receptor	1L
	2	Hypothalamic- hypophysial system and	15L
	0.4	Hormones of the adrenals	
	2.1	Hypothalamic - Pituitary axis: anatomy, histology, vasculature and secretions	2L
	2.2.1		
		hypothalamic hormones and Anterior pituitary	
		hormones	
	2.2.2	Hormone feed- back regulatory cascade	
als.	2.3.1	Posterior pituitary hormones –structure, physiology	3L
	2.3.2	Biochemical actions of AVP and Oxytocin;	
		Diabetes insipidus	
	2.4	Histology of Adrenal Gland	2L
	2.5	Physiology and action of Aldosterone; the Renin	
		Angiotensin System	
	2.6.1	Physiology and Biochemical actions of Cortisol	2L
	2.6.2	Regulation of cortisol synthesis: POMC and CRH	



	2.7.1	Adrenal medullary Hormones: Epinephrine and	2L
		Norepinephrine	
	2.7.2	The Fight or flight response; Dual receptor	
		hypothesis	
	2.8	General adaptation syndrome: acute and chronic	3L
		stress response	
	2.9	Pathophysiology – Addison's disease, Conn's	
		syndrome, Cushing syndrome.	
	3	Hormones regulating Metabolism, Calcium	15L
		homeostasis and Growth	CX
	3.1.1	Thyroid gland- Histology	3L
	3.1.2	Biosynthesis of thyroid hormone and its regulation:	
		Role of TRH and TSH in T4 synthesis and	
		response	
	3.1.3	Physiological and biochemical action of Thyroxine	3L
	3.1.4	Pathophysiology of thyroxine secretion: Hyper and	
		hypothyroidism, Goitre, Graves' disease,	
		Cretinism, Myxoedema	
	3.2.1	Regulation of calcium homeostasis: PTH, Vitamin	3L
III		D and calcitonin	
	3.2.2	Mechanism of Ca2+ regulation and pathways	
		involving bone, skin, liver, gut and kidneys	
	3.2.3	Pathophysiology - rickets, osteomalacia,	3L
		osteoporosis	
	3.3	Regulation of Growth: growth hormone and	
		somatomedin	
	3.4	Endocrine disorders - gigantism, acromegaly,	4L
		dwarfism, pygmies	
	3.5	Physiology and biochemical actions of Growth	
	1/1	factors- EGF, PDGF and EPO	
. 5	4	Pancreatic, GI tract and Reproductive	15L
	10.	hormones	
	4.1	Cells involved in release of gastrointestinal	3L
O ly		hormones	
	4.1.1	Gastrin family of hormones	
IV	4.1.2	CCK: the secretin family of hormones	
	4.1.3	Incretins; Ghrelin	
	4.2	Summary of hormone metabolite control of GI	2L
		function	
	4.3	Hormones of the Pancreas: Structure, synthesis,	
		physiology and biochemical actions of insulin and	
		glucagon	



4.4	Adipocyte hormones: Adiponectin and leptin;	3L
	Appetite and satiety control	
4.5	Pathophysiology Type I and type II Diabetes	
	mellitus, Obesity and Metabolic syndrome	
4.6	Male and female sex hormones	4L
4.6.1	Interplay of hormones during ovarian and uterine	
	phases of menstrual cycle	
4.6.2	Placental hormones; role of hormones during	
	parturition and lactation	
4.6.3	Hormone based contraception	3L
4.6.4	Concept - ammenorrhea, menorrhagia, PMS,	
	PCOS, Menopause	

	DDACTICALC	
	PRACTICALS	
RPSBCHP401	Project Work	4
&		Credits
RPSBCHP402		
	GUIDELINE TO CARRY OUT PROJECTWORK	
	1. The main purpose of introduction of Project Work at MSc	
	Il is to inculcate research culture. It will also make the	
	students familiar with Research Methodology i.e.	
	reference work, experimental work, data analysis of	
	experimental data, interpretation of results obtained,	
	writing of project work and compilation of bibliography in	
	proper order.	
	2. Each student shall complete a small research project	
	during their academic year of MSc II. However, the initial	
	reference work for the project can be started in MSc I.	
	3. Nature of Research Project:- Experimental-based or	
	literature survey involving laboratory analytical work will be	
	considered as the Research Project.	
	4. Duration of Project work:- Using the infrastructure	
	available in Ramnarain Ruia Autonomous College the	
	duration to complete the project work will be from the	
	commencement of the project work till the end of January	
	of MSc II (Sem IV) academic year.	
	The duration to complete the project work from any	
	institute apart from Ramnarain Ruia Autonomous College	
	will be 03 months (This needs prior approval from the	
	Department of Biochemistry, Ramnarain Ruia	
	Autonomous College).	
	5. Schedule for Submission of project Work:-	
	Experimental work or literature survey must be completed	



and the report on the same (2 Copies) will have to be submitted by the end of February of MSc II (Sem IV) academic year.

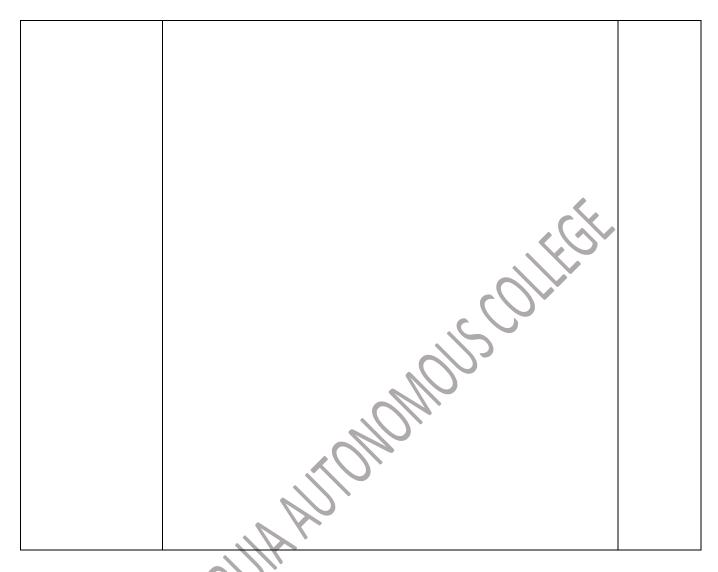
- 6. The project should be divided into the following parts:
 - a) Certification of completion of Project Work
 - b) Acknowledgement
 - c) Introduction
 - d) Review of Related Literature
 - e) Aims and Objectives
 - f) Plan of work
 - g) Material and Methods
 - h) Results
 - i) Discussion & Conclusion
 - j) Future Prospects
 - k) Bibliography
- 7. The project will be assessed.

GUIDELINE FOR THE ASSESMENT OF PROJECT WORK

- 1. The practical 401 & 402 of Sem IV (Course Code No. RPSBCHP401 & RPSBCHP402) shall be exclusively devoted for the project.
- Each student will complete the project (2 copies) and get both the copies certified by the guiding teacher and the Head of Dept. (HOD) by February of MSc II (Sem IV) academic year.
- 3. One copy of the certified project will be submitted to the Department; while the other copy will be retained by the students for his/ her personal record.
- 4. 4. The candidate is required to present the Research Project to the examiner followed by Viva- Voce examination based on the project work by the examiner.
- 5. The following Marking Scheme shall be considered while assessing the project work

	<u>Particular</u>	Marks
a)	Project Work (Contents	5
	Submitted in the bound form)	0
b)	Presentation of Project Work to	
	Examiner	3
c)	Viva- voce Exam based in	0
	Project Work	
		2
		0
	<u>TOTAL</u>	100





- Lehninger: Principles of Biochemistry (2013) 6th ed., Nelson, D.L. and Cox, M.M. W.H. Freeman & Company (NewYork), ISBN:13: 978-1-4641-0962-1 / ISBN:10-14641-0962-1.
- 2. Vander's Human Physiology (2008) 11th ed., Widmaier, E.P., Raff, H. and Strang, K.T. McGraw Hill International Publications, ISBN: 978-0-07-128366-3.
- 3. Endocrinology (2007) 6th ed., Hadley, M.C. and Levine, J.E. Pearson Education (New Delhi), Inc. ISBN: 978-81-317-2610-5.
- 4. Guyton and Hall Textbook of Medical Physiology 13th Edition by John E. Hall, Elsevier
- 5. Harrison's Endocrinology, Second Edition by J. Larry Jameson
- 6. The Cell: A Molecular Approach (2009) 5th Ed. Cooper, G.M. and Hausman, R.E. ASM Press & Sunderland, (Washington DC), Sinauer Associates. (MA). ISBN:978-0-87893- 300-6.



Course Code: RPSBCH403

Course Title: Genetic Engineering & Biotechnology

Academic year 2021-22

COURSE OUTCOMES:

COURSE OUTCOME	DESCRIPTION
CO 1	Acquire wide knowledge about Recombinant DNA technology by studying about various Vectors and Restriction Enzymes involved.
CO 2	Study of Various Expression Systems and Molecular Markers
CO 3	Apply the fundamental knowledge to understand Application of R-DNA technology and use of Restriction enzymes in construction of various vectors and libraries such as c-DNA & Genomic libraries



CO 4	Recognize the importance of Screening of the libraries with the help of "Reporter Genes" and Molecular Markers such as RFLP,RAPD, AFLP.
CO 5	Analyse and Interpret the knowledge about Genetic engineering and prospects of improving crop productivity, resistance, resistance to disease and environmental stresses, methods for production of transgenic animals.
CO 6	Develop innovative methods & Apply learned knowledge to their future research

prospects of improving crop productivity, resistance, resistance to disease and environmental stresses, methods for production of transgenic animals.							
CO 6		Develop innovative methods & Apply learned knowledge to their					
CO 6	future research						
	Tut	ure research					
		DETAILED SYLLABUS					
Course	Unit∜		Credits/				
Course Code/	Unit	Course/ Unit Title	Credits/				
Code/	Unit	Course/ Unit Title Genetic Engineering & Biotechnology	Lectures				
		Course/ Unit Title Genetic Engineering & Biotechnology RPSBCH403	Lectures 4 Credits				
Code/		Course/ Unit Title Genetic Engineering & Biotechnology RPSBCH403 Introduction to RDT & cloning vectors	Lectures 4 Credits 15L				
Code/		Course/ Unit Title Genetic Engineering & Biotechnology RPSBCH403 Introduction to RDT & cloning vectors Overview of RDT, Extraction and purification of	Lectures 4 Credits				
Code/		Course/ Unit Title Genetic Engineering & Biotechnology RPSBCH403 Introduction to RDT & cloning vectors Overview of RDT, Extraction and purification of plasmid and bacteriophage DNA	Lectures 4 Credits 15L				
Code/		Course/ Unit Title Genetic Engineering & Biotechnology RPSBCH403 Introduction to RDT & cloning vectors Overview of RDT, Extraction and purification of plasmid and bacteriophage DNA Restriction and modification systems, restriction	Lectures 4 Credits 15L				
Code/	1.1	Course/ Unit Title Genetic Engineering & Biotechnology RPSBCH403 Introduction to RDT & cloning vectors Overview of RDT, Extraction and purification of plasmid and bacteriophage DNA Restriction and modification systems, restriction endonucleases, Concept of sticky ends, blunt ends	Lectures 4 Credits 15L 3L				
Code/		Course/ Unit Title Genetic Engineering & Biotechnology RPSBCH403 Introduction to RDT & cloning vectors Overview of RDT, Extraction and purification of plasmid and bacteriophage DNA Restriction and modification systems, restriction endonucleases, Concept of sticky ends, blunt ends Other enzymes used in manipulating DNA	Lectures 4 Credits 15L				
Code/	1.1	Course/ Unit Title Genetic Engineering & Biotechnology RPSBCH403 Introduction to RDT & cloning vectors Overview of RDT, Extraction and purification of plasmid and bacteriophage DNA Restriction and modification systems, restriction endonucleases, Concept of sticky ends, blunt ends Other enzymes used in manipulating DNA molecules:	Lectures 4 Credits 15L 3L				
Code/	1.1	Course/ Unit Title Genetic Engineering & Biotechnology RPSBCH403 Introduction to RDT & cloning vectors Overview of RDT, Extraction and purification of plasmid and bacteriophage DNA Restriction and modification systems, restriction endonucleases, Concept of sticky ends, blunt ends Other enzymes used in manipulating DNA molecules: Terminal transferases, linkers and adapters,	Lectures 4 Credits 15L 3L				
Code/	1.1	Course/ Unit Title Genetic Engineering & Biotechnology RPSBCH403 Introduction to RDT & cloning vectors Overview of RDT, Extraction and purification of plasmid and bacteriophage DNA Restriction and modification systems, restriction endonucleases, Concept of sticky ends, blunt ends Other enzymes used in manipulating DNA molecules:	Lectures 4 Credits 15L 3L				



	1.3.2	Synthetic oligonucleotides - synthesis and use	2L		
	1.3.3	Plasmids and bacteriophages as vectors for gene	4L		
		cloning			
	1.4	Cloning vectors based on E. coli plasmids,			
		pBR322, pUC8, pGEM3Z			
	1.5	Cloning vectors based on M13 and λ			
		bacteriophage, and in vitro packaging Vectors for			
		yeast, Ti-plasmid, and retroviral vectors, high			
		capacity vectors			
	1.5.1 BAC and YAC				
	2	Gene transfer processes, selection for	2L 15L		
		recombinants & clone identification	X >		
	2.1	Transformation	3L		
	2.1.1	Preparation of competent cells using			
		transformation and selection for transformed cells			
	2.1.2	Identification for recombinants - insertional			
		inactivation, blue-white selection Conjugation (F+&			
		F- cells)			
	2.2.1	Mechanism of conjugation in Hfr strains	3L		
	2.2.2	Interrupted mating experiment technique			
	2.2.3	Problems based on above concept			
	2.2.4	Transduction (Generalized & Specialized)			
II	2.3	Introduction of phage DNA into bacterial cells	2L		
	2.3.1	Identification of recombinant phages			
	2.3.2	Transfection			
	2.4	Transient and stable transfection	5L		
	2.4.1	Chemical and physical methods of transfection			
	2.4.2	The problem of selection, direct selection, marker			
	-	rescue.			
	2.4.3	cDNA and Genomic libraries, identification of a			
		clone from gene library, colony and plaque			
	171,	hybridization probing, Southern and Northern			
		hybridization			
	2.5	Methods based on detection of the translation	2L		
		product of the cloned gene			
	3	Expression of cloned genes, PCR & DNA	15L		
		sequencing			
	3.1.1	Vectors for expression of foreign genes in E. coli,	3L		
III		cassettes and gene fusions			
	3.1.2	Challenges in producing recombinant protein in E.			
	0.0	coli	4:		
	3.2	Production of recombinant protein by eukaryotic	4L		
		cells			



	3.2.1	Fusion tags such as, poly-histidine, glutathione, maltose binding protein and their role in purification of recombinant proteins	
	3.3.1	Fundamentals of polymerase chain reaction	5L
	3.3.2	Types of PCR – hot start, multiplex, reverse transcriptase PCR and Nested PCR, quantitative PCR, Primer, designing for PCR, Cloning PCR products	
	3.4	DNA sequencing by Sanger's method, Automated Sanger's DNA sequencing, Pyrosequencing	3L
	4	Application of genetic engineering in Biotechnology	15L
	4.1	Site-directed mutagenesis (original method, Kunkel's method, cassette mutagenesis, PCR oligonucleotide mutagenesis), Protein engineering (T4-lysozyme), yeast two hybrid systems	4L
	4.2	Production of recombinant pharmaceuticals such as insulin, human growth hormone (original, receptor fragment-hormone coupled, albutropin), factor VIII.	2L
IV	4.3	Recombinant vaccines	1L
	4.4	Gene therapy & its application; CRISPR-Cas 9 system	1L
	4.5	Applications in agriculture – Bt cotton, problems with genetically modified plants, glyphosate herbicide resistant crops, ethical & safety concerns	3L
	4.6	RDT in diagnosis and treatment of diseases	2L
	4.7	Model organisms: Escherichia coli, Saccharomyces cerevisiae, Drosophila melanogaster, Caenorhabditis elegans, Danio rerio	2L
	16	and Arabidopsis thaliana	
	1 2	Practicals – RPSBCHP403 Isolation of plasmid DNA from E. coli cells Separation of chromosomal & plasmid DNA using	2 Credits
KL	3 4 5	agarose gel electrophoresis Digestion of plasmid DNA with restriction enzymes Preparation of competent cells (CaCl ₂ treatment) Transformation of E. coli cells with plasmid DNA	
	6	Demonstration of complementation of	
	7	β – galactosidase for Blue and White selection	
		β-galactosidase Activity Assay in Permeabilized Yeast	



- 1. Gene Cloning and DNA Analysis (2010) 6th ed., Brown, T.A., Wiley-Blackwell publishing (Oxford, UK), ISBN: 978-1-4051-8173-0.
- 2. Principles of Gene Manipulation and Genomics (2006) 7th ed., Primrose, S.B., and Twyman, R. M., Blackwell publishing (Oxford, UK) ISBN:13: 978-1-4051-3544-3.
- 3. Molecular Biotechnology: Principles and Applications of Recombinant DNA (2010) 4th ed., Glick B.R., Pasternak, J.J. and Patten, C.L., ASM Press (Washington DC), ISBN: 978-1-55581-498-4 (HC).
- 4. Genetic engineering and its applications, P. Joshi, Botania Publishers and Distributors
- 5. Recombinant DNA: A short course, Watson etal, Scientific Americal Books
- 6. Biotechnology Fundamentals and Applications, S.S.Purohitt, Agrobios Publishers, 2001.
- 7. Molecular Biology of the Gene: Watson, Baker, Bell, Gann, Levine, Losick; Pearson Benjamin Cummings & CSHL Press
- 8. Gene cloning & DNA analysis: an introduction; seventh edition; T A Brown; Wiley Blackwell publications

Course Code: RPSBCH404

Course Title: Advanced immunology

Academic year 2021-22

COURSE OUTCOMES:

COURSE OUTCOME	DESCRIPTION
CO 1	Learn the fundamental principles of immune response including molecular, biochemical and cellular basis of immune homeostasis
CO 2	Develop various aspects of immunological response and how its triggered and regulated



CO 3	Explain the specific interactions of Antigens and antibodies and the
	diversity of antibodies developed at the germ line DNA
CO 4	Complete knowledge of the molecular mechanisms and kinetics of
	the immune responses, both humoral and cell mediated immunity
CO 5	Enhance the knowledge of various immune-techniques ranging
	from precipitation and agglutination reactions to ELISA, Radio
	immunoassay and flow cytometry.
CO 6	The course will aid in understanding the principles of Graft
	rejection, Auto immunity and Antibody based therapy.

CO 4	Co	emplete knowledge of the molecular mechanisms and	d kinetics of					
	the	e immune responses, both humoral and cell mediated	immunity					
CO 5	Er	hance the knowledge of various immune-techniques	ranging					
	fro	om precipitation and agglutination reactions to ELISA,	Radio					
	im	munoassay and flow cytometry.						
CO 6 The course will aid in understanding the principles of Graft								
	rejection, Auto immunity and Antibody based therapy.							
Course								
Course	Unit	Course/ Unit Title	Credits/					
Code/	17,	Advanced immunology	Lectures					
Unit		RPSBCH404	4 Credits					
Bly	1	Antigen-Antibody Interactions and Complement System	15L					
	1.1.1	Overview of Antigens and Antibodies	1L					
	1.1.2	Antigen- Antibody interactions: Forces involved,	2L					
I		antibody affinity, antibody avidity, Cross reactivity						
	1.1.3	Precipitation reactions – Oudins, Ouchterlony	2L					
	1.1.4	Agglutination reactions: Blood typing, bacterial						
		agglutination, passive agglutination, agglutination inhibition,						



	1		
	1.1.5	Immunoelectrophoresis: Principles of	3L
		Radioimmunoassay, ELISA, Immunofluorescence,	
		Western Blotting	
	1.2	Complement system	
	1.2.1	Function and components	2L
	1.2.2	Complement activation - Classical, Alternate &	3L
		Lectin pathways.	
	1.2.3	Biological consequence of complement activation –	3L
		formation of MAC	
	2	MHC, Antigen processing and presentation & TCR	15L
	2.1	Major histocompatibility complex	
	2.1.1	MHC polymorphism & organization of MHC genes-	O.I.
		class I & class II	2L
	2.1.2	Cellular distribution & structure of class & II	
		molecules	
	2.1.3	MHC and immune responsiveness - Determinant-	2L
		selection model and Holes-in-the-repertoire model	
	2.1.4	MHC and disease susceptibility (Hereditary	2L
II		haemochromatosis)	
	2.2	Antigen processing and presentation	4L
	2.2.1	Self MHC restriction of T cells	
	2.2.2	Cytosolic and endocytic pathway	
	2.2.3	Presentation of non-peptide antigens	
	2.3	T-cell Receptor	2L
	2.3.1	Structure, organization & rearrangement of TCR genes	
	2.3.2		1L
	2.3.3	TCR accessory membrane molecules	1L
	2.3.4		1L
	3	B- and T-lymphocytes	15L
	3.1	T-cell Maturation	2L
Un,	3.2	Thymic selection of T-cell repertoire – Positive and	3L
Olyn	0.2	negative selection, central issues in thymic	02
161		selection	
	3.3	TH-cell activation	
III	3.4	T-cell differentiation	1L
	3.5	Peripheral γδ T-cell	2L
	3.6	Cytotoxic T-cells	2L
	3.7	B-cell maturation	4L
	3.8	B-cell activation and proliferation – Thymus	⊤∟
	0.0	dependent and Thymus independent	
		aoponaoni ana myinao maepenaem	



	3.9	Formation of T-B conjugates	1L
	4	Cytokines & Immune response in health & diseases	15L
	4.1	Humoral and cell mediated immune response	2L
	4.1.1	Cytokines - Introduction, Properties	
	4.1.2	Cytokine receptors	3L
	4.1.3	Biological functions of cytokines	
	4.1.4	Therapies based on Cytokines	
	4.2	Hypersensitivity reactions	3L
IV	4.2.1	Gel & Coomb's classification - types of	. (.)
1 1 4		hypersensitivity reactions	
	4.3	Transplantation immunology	3L
	4.3.1	Types of transplant; immunological basis of	
		allograft rejection.	
	4.4	Autoimmunity	4L
	4.4.1	Organ specific – Myasthenia gravis; Hashimotos	
		thyroiditis; Graves' Disease; Systemic –	
		Rheumatoid arthritis, Systemic lupus	
		erythematosus	
		Practicals – RPSBCHP404	2 Credits
	1	Preparation of blood smear and Differential	
	_	leucocyte count.	
	2	Separation of lymphocytes by Ficoll Hypaque	
		method	
	3	Lymphocyte viability testing by trypan blue	
	4	Assays based on precipitation reactions -	
		Ouchterlony double immunodiffusion (DID) and	
		Mancini radial immunodiffusion (SRID).	
		Assays based on agglutination reactions - Blood	
	2	typing (active) & passive agglutination (C reactive protein kit & virtual lab).	
	MIL	Demonstration of Enzyme linked immunosorbent	
	6	assay (ELISA) & DOT ELISA	
		Separation of serum proteins by PAGE	
17/L	7	Virtual Lab to study immunological Techniques	
	8	This are to stay in anotogical reciniques	

- 1. Kuby Immunology (2007) 6th ed., Kindt, T.L., Goldsby, R.A. and Osborne, B.A., W.H Freeman and Company (New York), ISBN:13: 978-0-7167-8590-3 / ISBN: 10:0-7617-8590-0.
- 2. Immunology: A Short Course (2009) 6th ed., Coico, R and Sunshine, G.,



- John Wiley& sons, Inc (New Jersey), ISBN: 978-0-470-08158-7.
- 3. Janeway's Immunobiology (2012) 8th ed., Murphy, K., Mowat, A., and Weaver, C.T., Garland Science (London & New York), ISBN: 978-0-8153-4243-4.
- 4. Willey, J. Sherwood L, Woolverton C, (2016), Prescott Microbiology. 10th Edition, McGraw-Hill Publisher, Columbus, OH
- ARINIARINA RINARIA RIN 5. Pelczar Mi J., Chan, E.C.S., Krieg, NR, (2009). Microbiology, McGraw-Hill



Modality of Assessment (SEMESTER IV)

Theory Examination Pattern:

- A) Internal Assessment- 40%- 40 Marks
- B) External Examination- 60%- 60 Marks
 Semester End Theory Examination: (Deviation from the usual modality)
 Owing to the pandemic situation prevailing in 2020 and continuing in 2021, the external examinations (Semester End) may be conducted online as per the instructions/circulars received from the University of Mumbai and Maharashtra State notifications from time to time. The conventional mode of external examination will commence again only after the declaration of normalcy by the Government authorities.
 - 1. Duration These examinations shall be of 02 ½ HOURS duration.
 - 2. Theory question paper pattern:

Paper Pattern:

Question	Options	Marks	Questions Based on
Q1. A	Any 1 out of 2	03	LINUT
Q1. B	Any 2 out of 3	06	UNIT I
Q2. A	Any 1 out of 2	03	UNIT II
Q2. B	Any 2 out of 3	06	UNIT II
Q3. A	Any 1 out of 2	03	UNIT III
Q3. B	Any 2 out of 3	06	ONIT III
Q4. A	Any 1 out of 2	03	UNIT IV
Q4. B	Any 2 out of 3	06	OINIT IV
	TOTAL	60	



Practical Examination Pattern:

A) Internal Examination: 40%-40 Marks

Particulars	Practical I, II, III & IV
Journal	05
Experimental	15
tasks	
Total	20

B) External Examination: 60%-60 Marks

Semester End Practical Examination:

Particulars	Practical I, II, III & IV
Laboratory work	25
Viva	5
Total	30

Overall Examination & Marks Distribution Pattern

Semester IV

Course 401				4	02		Grand Total
	External	Total	Internal	External	Total		
Theory	40	60	100	40	60	100	200
Practicals	20	30	50	20	30	50	100

Course	403			404			Grand Total
	Internal	External	Total	Internal	External	Total	
Theory	40	60	100	40	60	100	200
Practicals	20	30	50	20	30	50	100
