

**S. P. Mandali's**  
**Ramnarain Ruia Autonomous College**  
*(Affiliated to University of Mumbai)*



**Syllabus for**

**Program: MSc Part II**

**Program Code: RPSBCH**

(Credit Based Semester and Grading  
System for academic year 2020–2021)

## PROGRAM OUTCOMES

PO	PO Description
	<b>A student completing Master's Degree in SCIENCE program will be able to:</b>
PO 1	Demonstrate in depth understanding in the relevant science discipline. Recall, explain, extrapolate, and organize conceptual scientific knowledge for execution and application and also to evaluate its relevance.
PO 2	Critically evaluate, analyse, and comprehend a scientific problem. Think creatively, experiment and generate a solution independently, check and validate it and modify if necessary.
PO 3	Access, evaluate, understand, and compare digital information from various sources and apply it for scientific knowledge acquisition as well as scientific data analysis and presentation.
PO 4	Articulate scientific ideas, put forth a hypothesis, design and execute testing tools and draw relevant inferences. Communicate the research work in appropriate scientific language.
PO 5	Demonstrate initiative, competence, and tenacity at the workplace. Successfully plan and execute tasks independently as well as with team members. Effectively communicate and present complex information accurately and appropriately to different groups.
PO 6	Use an objective, unbiased and non-manipulative approach in collection and interpretation of scientific data and avoid plagiarism and violation of Intellectual Property Rights. Appreciate and be sensitive to environmental and sustainability issues and understand its scientific significance and global relevance.
PO 7	Translate academic research into innovation and creatively design scientific solutions to problems. Exemplify project plans, use management skills, and lead a team for planning and execution of a task.
PO 8	Understand cross disciplinary relevance of scientific developments and relearn and reskill so as to adapt to technological advancements.

## PROGRAM SPECIFIC OUTCOMES

PSO	Description
	<b>A student completing Master's Degree in SCIENCE program in the subject of BIOCHEMISTRY will be able to:</b>
PSO 1	Acquire necessary knowledge and skills to undertake a career in research, either in industry or in an academic set up.
PSO 2	Compare and contrast the breadth and depth of scientific knowledge in the broad range of fields including Protein biochemistry, Bioenergetics, Diagnostic Biochemistry, Hormonal Biochemistry, Molecular Biology, Nutritional Biochemistry, and Nanotechnology.
PSO 3	Extrapolate and comprehend the regulatory role of metabolic processes and understand the underlying cause of metabolic disorders
PSO 4	Acquire thorough knowledge of Biochemical Techniques, Advanced Immunology, Physiology, Genetic Engineering, and Biotechnology
PSO 5	Describe and express the biochemical basis of human diseases, protein structure and conformation, non-invasive diagnostics, clinical research, and its importance in drug development. Usage of this knowledge further for multitude of laboratory applications.
PSO 6	Integrate and apply the techniques in Biophysics, Analytical Biochemistry, Clinical biochemistry, Microbiology, Molecular Biology and Basics in Bioinformatics
PSO 7	Gain proficiency in laboratory techniques in both Biochemistry and Molecular Biology, and be able to apply the scientific method to the processes of experimentation and Hypothesis testing
PSO 8	Develop and enhance skills & improve employability through academic, research and internship opportunities
PSO 9	Gain exposure to basic research through the provision of PG research based project.
PSO 10	Learn to work as a team as well as independently to compile and interpret Biological data, carry out Research investigations and draw conclusions

## PROGRAM OUTLINE

YEAR	SEM	COURSE CODE	COURSE TITLE	CREDITS
MSc II	III	RPSBCH301	Biochemistry of Metabolism	4
		RPSBCH302	Clinical biochemistry	4
		RPSBCH303	Molecular biology	4
		RPSBCH304	Biology of diseases & Clinical Research	4
		RPSBCHP301	Practicals based on RPSBCH301	2
		RPSBCHP302	Practicals based on RPSBCH302	2
		RPSBCHP303	Practicals based on RPSBCH303	2
		RPSBCHP304	Practicals based on RPSBCH304	2
	IV	RPSBCH401	Nutritional Biochemistry	4
		RPSBCH402	Endocrinology	4
		RPSBCH403	Genetic Engineering & Biotechnology	4
		RPSBCH404	Advanced Immunology	4
		RPSBCHP401	Dissertation	4
		RPSBCHP402		
		RPSBCHP403	Practicals based on RPSBCH403	2
		RPSBCHP404	Practicals based on RPSBCH404	2

**Course Code: RPSBCH301****Course Title: Biochemistry of Metabolism****Academic year 2020-21****COURSE OUTCOMES:****After completion of the course, a student will be able to achieve these outcomes**

<b>COURSE OUTCOME</b>	<b>DESCRIPTION</b>
<b>CO 1</b>	Understand the major catabolic and anabolic pathways in metabolism of carbohydrates, lipids, amino acids and nucleotides.
<b>CO 2</b>	Describe regulatory mechanisms that control the metabolic pathways.
<b>CO 3</b>	Realize the Influence of Diet and hormonal signalling on metabolic pathways.
<b>CO 4</b>	Learn Biochemical functions and integrated metabolism of in brain, digestive system, liver, red cell, muscle and adipocyte.
<b>CO 5</b>	Illustrate the molecular mechanisms underlying major inherited diseases of metabolism.
<b>CO 6</b>	Understand the relationship between the properties of macromolecules and cellular activities, cell metabolism and chemical composition.

## DETAILED SYLLABUS

Course Code/ Unit	Unit	Course/ Unit Title Biochemistry of Metabolism RPSBCH301	Credits/ Lectures 4 Credits
<b>I</b>	<b>1</b>	<b>Carbohydrate Metabolism</b>	<b>15L</b>
	1.1	Schematic representation of Glycolysis & Kreb's cycle	2L
	1.2	Gluconeogenesis: Pathway and its Regulation	1L
	1.3	Glycogen Metabolism: Synthesis, breakdown, mechanisms of control of glycogen metabolism - Direct Allosteric Control of Glycogen Phosphorylase and Glycogen Synthase, Covalent Modification of Enzymes by Cyclic Cascades, Hormonal regulation, Maintenance of Blood Glucose Levels	3L
	1.4	Futile cycle, Rapoport Luebering cycle, Cori cycle, Glucose-Alanine cycle & their significance	2L
	1.5	Shuttles-Malate-Aspartate shuttle & Glycerol phosphate shuttle	3L
	1.6	Uronic acid pathway (biosynthesis, degradation & its significance), Galactose and fructose metabolism; Sorbitol pathway	
	1.7	Biosynthesis of oligosaccharides and glycoproteins	1L
	1.8	Synthesis of carbohydrates in plants	3L
	1.8.1	Calvin cycle and its regulation, regulated synthesis of starch and sucrose, photorespiration, C4 and CAM pathways, Glyoxylate pathway, synthesis of cell wall polysaccharides	
<b>II</b>	<b>2</b>	<b>Amino acid metabolism</b>	<b>15L</b>
	2.1	Reactions of amino acids: Deamination, Transamination, Decarboxylation, Transmethylation, Transdeamination,	3L
	2.2	Ammonia formation, transport and detoxification in brain and liver	
	2.3	Kreb's bicycle, Urea cycle & its regulation. Inherited defects of urea cycle	2L
	2.4	Glucogenic and ketogenic amino acids.	
2.5	Metabolism of one carbon units. Disorders of amino acids metabolism, phenylketonuria, alkaptonuria, maple syrup urine disease,	3L	

		methylnmalonic acidemia (MMA), homocystinuria and Hartnup's disease	
	2.6	Biosynthesis of amino acids Overview of amino acid synthesis. Biosynthesis of non-essential amino acids and its regulation	2L
	2.7	Precursor functions of amino acids	1L
	2.7.1	Biosynthesis of creatine and creatinine, polyamines (putresine, spermine, spermidine), catecholamines (dopamine, epinephrine, norepinephrine) and neurotransmitters (serotonin, GABA).	4L
III	<b>3</b>	<b>Lipid metabolism</b>	<b>15L</b>
	3.1	Fatty acid oxidation	4L
	3.1.1	Fatty acid transport to mitochondria, $\beta$ -oxidation of saturated, unsaturated, odd and even numbered and branched chain fatty acids, regulation of fatty acid oxidation, peroxisomal $\beta$ -oxidation, $\omega$ oxidation, ketone bodies metabolism, ketoacidosis	
	3.2	Fatty acid synthesis	2L
	3.2.1	Transport of mitochondrial Acetyl Co A to cytosol, Fatty acid synthase complex, Synthesis of saturated, unsaturated, odd and even chain fatty acids and regulation.	
	3.2.2	Biosynthesis of eicosanoids, cholesterol, steroids and isoprenoids	2L
	3.2.3	Synthesis of prostaglandins, leukotrienes and thromboxanes. Synthesis of cholesterol, regulation of cholesterol synthesis. Synthesis of steroids and isoprenoids.	3L
	3.2.3	Biosynthesis of glycerophospholipids and sphingolipids	2L
	3.4	Lipid storage diseases	2L
IV	<b>4</b>	<b>Nucleic Acid Metabolism &amp; Integration of Metabolism</b>	<b>15L</b>
	4.1	Biosynthesis of purine and pyrimidine nucleotides	3L
	4.1.1	De novo synthesis of purine and pyrimidine nucleotides, regulation and salvage pathways. Deoxyribonucleotides and synthesis of nucleotide triphosphate	
	4.1.2	Biosynthesis of deoxyribonucleotides and its regulation, conversion to triphosphates,	2L
	4.2	Biosynthesis of coenzyme nucleotides	1L
	4.3	Degradation of purine and pyrimidine nucleotides	2L

	4.4	Digestion of nucleic acids, degradation of purine and pyrimidine nucleotides. Inhibitors of nucleotide metabolism.	1L
	4.5	Disorders of purine and pyrimidine metabolism – Lesch-Nyhan, syndrome, Gout, SCID, adenosine deaminase deficiency	3L
	4.6	Integration of metabolism	1L
	4.6.1	Integration of metabolic pathways (carbohydrate, lipid and amino acid metabolic pathways), tissue specific metabolism (brain, muscle, and liver).	2L
		<b>Practicals – RPSBCHP301</b>	2 Credits
	1	Estimation of glucose by the Folin-Wu method	
	2	Estimation of glucose by the GOD-POD method	
	3	Demonstration of glucose metabolism using handheld glucometer	
	4	Assay of serum transaminases – SGOT and SGPT.	
	5	Estimation of serum urea.	
	6	Estimation of serum creatinine.	
	7	Assay of glutamate dehydrogenase	
	8	Lipid Profile – a) Estimation of total cholesterol and HDL b) Estimation of Triglycerides c) Estimation of LDL by calculation	
	9	Estimation of serum uric acid by phosphotungstic acid method (Caraways method)	
	10	Use of softwares to understand metabolism – KEGG, Ecocyc, Metacyc, Biocyc	

**References:**

1. Lehninger: Principles of Biochemistry (2013) 6th ed., Nelson, D.L. and Cox, M.M., W.H. Freeman and Company (New York), ISBN:13:978-1-4641-0962-1 / ISBN:10:1-4641-0962-1.
2. Textbook of Biochemistry with Clinical Correlations (2011) 7th ed., Devlin, T.M., John Wiley & Sons, Inc. (New Jersey), ISBN:978-0-470-28173-4.
3. Biochemistry (2012) 7th ed., Berg, J.M., Tymoczko, J.L. and Stryer L., W.H. Freeman and Company (New York), ISBN:10:1-4292-2936-5, ISBN:13:978-1-4292-2936-4.
4. Principles of Biochemistry by G. Zubay, W. Parson, D.
5. Biochemistry - Voet, D. and Voet, J.G. - John Wiley & Sons, Inc. USA.



**Course Code: RPSBCH302**  
**Course Title: Clinical Biochemistry**  
**Academic year 2020-21**

**COURSE OUTCOMES:**

After completion of the course, a student will be able to achieve these outcomes

<b>COURSE OUTCOME</b>	<b>DESCRIPTION</b>
<b>CO 1</b>	The student will learn and understand the basics of circulatory system including haematopoiesis, homeostasis, and diseases of blood.
<b>CO 2</b>	Explain the fundamentals, composition and significance of Body fluids
<b>CO 3</b>	To get acquainted with the role of enzymes in diagnosis of various diseases.
<b>CO 4</b>	The student will be aware of the organ function tests available for detection of their functionality
<b>CO 5</b>	Illustrate Pathophysiology of certain disorders related to certain organs & Clinical tests available for detection

## DETAILED SYLLABUS

Course Code/ Unit	Unit	Course/ Unit Title Clinical Biochemistry RPSBCH302	Credits/ Lectures 4 Credits
<b>I</b>	<b>1</b>	<b>Haematopoiesis &amp; related disorders</b>	<b>15L</b>
	1.1	Introduction to Haematopoiesis	1L
	1.1.1	Erythropoiesis - Stages of development of erythrocytes, Precursors of RBCs, Factors influencing erythropoiesis	2L
	1.1.2	Role of erythropoietin	
	1.2	Haemoglobin (Hb)-Features, varieties, combination of Hb with gases, Haeme-haeme interactions	4L
	1.2.1	Biosynthesis of Haemoglobin (with structures)	
	1.2.3	Biochemical pathway for Porphyrin synthesis, formation of Haeme	
	1.3	Haeme catabolism	2L
	1.4	Haemoglobinopathies	2L
	1.4.1	Genetics basis of haemoglobinopathies - Sickle cell anemia, Thalassemia – alpha (Subtypes of alpha thalassemia) & beta	
	1.5	Blood gas analysis	1L
	1.5.1	Leucopoiesis, Leucocytosis and factors responsible, Leukopenia, Thrombopoiesis, Thrombocytopenia	2L
		1.6	Molecular mechanism of blood coagulation, role of vitamin K in coagulation, anticoagulant and fibrinolytic systems, haemophilia and thrombosis
<b>II</b>	<b>2</b>	<b>Body Fluids</b>	<b>15L</b>
	<b>2.1</b>	<b>Plasma</b>	<b>3L</b>
	2.1.1	Composition, Separation of plasma proteins – Salting out, Cohn's fractionation, Electrophoresis	
	2.1.3	Characteristics, functions and clinical significance of plasma proteins	
	2.2	Proteinuria – types & causes	1L
	2.3	Bile	4L
	2.3.1	Bile pigments- Biliverdin and Bilirubin	
	2.3.2	(formation, transport, conjugation in liver cells, secretion in bile, excretion)	
	2.3.3	Clinical importance of bile formation (concept - index of rate of haeme catabolism, shunt hyperbilirubinaemia, bilirubin encephalopathy)	

	2.4	Lymph- Formation, composition and circulation	1L
	2.5	CSF- Composition, Appearance, Pressure of CSF and Biochemical Changes in CSF and its clinical significance	2L
	2.6	Synovial fluid- Composition, Classification and Clinical significance – synovial fluid viscosity, pathology	1L
	2.6.1	Analysis – mucin clot test, microscopic analysis	1L
	2.7	Pericardial fluid- Composition, Function & Pericardiocentesis, Pericardial effusion, Ischemic heart disease	2L
	<b>3</b>	<b>Enzymes &amp; isoenzymes of clinical importance</b>	<b>15L</b>
<b>III</b>	3.1	Introduction, Possible mechanisms responsible for abnormal enzyme levels	4L
	3.2	Value of serum enzyme assay in clinical practice Enzyme assays of clinical significance	
	3.3	Myocardial infarction – Creatine phosphokinase, S-GOT, LDH. Important cardiac markers useful in diagnosis of acute myocardial infarctions	4L
	3.4	GI tract diseases - Serum amylase, serum lipase	2L
	3.5	Liver diseases – Serum transaminases	
	3.6	Muscle diseases – Serum aldolase, serum CPK	2L
	3.7	Bone diseases – Serum alkaline phosphatases	
	3.8	Malignancies – Acid phosphatase, $\beta$ -glucuronidase	1L
	3.9	Value & clinical significance of isoenzymes of – LDH, CPK, Alkaline phosphatase	2L
		<b>4</b>	<b>Organ Function Tests</b>
<b>IV</b>	4.1	Renal Function test Preliminary investigations Classification of renal function tests – Tests based on glomerular filtration, Tests to measure Renal Plasma Flow, Tests based on tubular function	2L
	4.2	Liver Function test Tests based on abnormalities of bile pigment metabolism - VD Bergh Reaction, Determination of Serum Bilirubin Tests based on liver's part in carbohydrate metabolism – Glucose tolerance test, Galactose tolerance test, Fructose tolerance test	2L
	4.3	Tests based on changes in plasma proteins – Determination of Total Plasma Proteins and Albumin and Globulin and A:G Ratio, Estimation of Plasma Fibrinogen, Flocculation Tests, Amino acids in urine	3L

	4.4	Tests based on abnormalities of lipids - Determination of serum cholesterol and ester cholesterol and their ratio, Determination of faecal fats	2L
	4.5	Tests based on detoxicating function of liver - Hippuric acid synthesis test	2L
	4.6	Gastric Function test – Fractional gastric analysis, stimulation tests	2L
	4.7	Thyroid Function test Tests based on primary function of thyroid, Tests measuring blood levels of thyroid hormones	2L
		<b>Practicals – RPSBCHP302</b>	2 Credits
	1	Enumeration of Blood cells: RBC and WBC counting	
	2	Qualitative analysis of bile	
	3	Estimation of serum electrolytes	
	4	Estimation of blood glucose	
	5	Estimation of serum phosphorus by Fiske Subbarow method	
	6	Estimation of serum Calcium	
	7	Clinical analysis of CSF – glucose, proteins, chlorides	
	8	Liver Function Tests – a) Estimation of serum ALT and AST b) Estimation of total and direct bilirubin c) Estimation of serum alkaline phosphatase d) Estimation of total proteins, albumin and determination of A/G ratio e) Estimation of serum albumin by Bromocresol Green (BCG) binding method	
	9	Renal Function tests a) Creatinine clearance test b) Urea clearance test	
	10	Pancreatic Function Test a) Estimation of serum amylase activity b) Glucose Tolerance Test	
	11	Estimation of serum total cholesterol and HDL cholesterol	
	12	Estimation of triglycerides	

**References:**

1. Textbook of Biochemistry with Clinical Correlations (2011) Devlin, T.M. John Wiley & Sons, Inc. (New York), ISBN: 978-0-4710-28173-4.

2. Introduction to Human Physiology (2013) 8th edition; Lauralee Sherwood. Brooks/Cole, Cengage Learning.
3. The World of the cell, 7th edition (2009)
4. Genetics (2012) Snustad and Simmons
5. Clinical Biochemistry Metabolic and Clinical Aspects by William J. Marshall, Márta Lapsley, Andrew Day, Ruth Ayling
6. Cooper, G.M. and Hausman, R.E. 2009 The Cell: A Molecular Approach. 5th edition. ASM Press & Sunderland, Washington, D.C.; Sinauer Associates, MA Principles of Biochemistry by G. Zubay, W. Parson, D.

RAMNARAIN RUIA AUTONOMOUS COLLEGE

**Course Code: RPSBCH303**  
**Course Title: Molecular biology**  
**Academic year 2020-21**

**COURSE OUTCOMES:**

After completion of the course, a student will be able to achieve these outcomes

<b>COURSE OUTCOME</b>	<b>DESCRIPTION</b>
<b>CO 1</b>	Understand the mechanism of cell cycle, relationship of cell cycle and programmed cell death via intracellular and extracellular control mechanisms
<b>CO 2</b>	The students will learn about nucleic acid as genetic information carriers, Possible modes of replication, and roles of helicase, primase, gyrase, topoisomerase, DNA Polymerase, DNA ligase, and Regulation of replication.
<b>CO 3</b>	The student will be able to illustrate the mechanism of prokaryotic and eukaryotic replications
<b>CO 4</b>	The student will learn & understand different types of mutations, agents causing mutations and disorders resulting from mutations.
<b>CO 5</b>	Comprehend the mechanism and regulation of transcription in prokaryotes along with Reverse transcription.
<b>CO 6</b>	The student will be able to describe synthesis of protein from gene with the help of regulatory protein
<b>CO 7</b>	The student will be able to explain the Post transcriptional & translational modifications & their significance in stability

## DETAILED SYLLABUS

Course Code/ Unit	Unit	Course/ Unit Title Molecular Biology RPSBCH303	Credits/ Lectures 4 Credits
I	<b>1</b>	<b>Cell Cycle and its regulation &amp; DNA Replication</b>	<b>15L</b>
	1.1	Cell cycle and its regulation	4L
	1.1.1	Phases of cell cycle and its regulation (Cyclins & CDKs)	
	1.1.2	State of DNA in different phases of cell cycle Replication of DNA	
	1.2	Structural overview of DNA Replication	5L
	1.2.1	Models for DNA replication-	
	1.2.2	Conservative, Semi-conservative & dispersive	
	1.2.3	Experimental evidences	
	1.2.4	Enzymes and proteins involved in replication	
	1.2.5	Mechanism of Bacterial DNA replication	
	1.2.6	Replication of DNA in yeast	
	1.3	Eukaryotic DNA polymerases	5L
	1.3.1	Proteins and accessory molecules essential in the initiation, and elongation steps	
	1.3.2	Mechanism (Pre-RC assembly, Initiation, elongation & termination)	
	1.3.3	Concept of Okazaki fragment maturation & stalled replication fork	
	1.3.4	End replication problem and role of telomerases	
	1.4	Comparative overview of DNA replication in prokaryotes and eukaryotes	1L
II	<b>2</b>	<b>Mutations, Chromosomal Abnormalities &amp; DNA Repair</b>	<b>15L</b>
	2.1	Mutations	3L
	2.1.1	Types of mutations Physical, chemical and Biological agents causing mutations	
	2.1.2	Reverse mutations, Mutagenesis, Ames test. Chromosomal aberration	
	2.1.3	Variations in chromosome structure - inversions, deletions, duplications and translocations	2L
	2.1.4	Variations in chromosome number - Euploidy and aneuploidy (Autosomal and Sex chromosomes)	
	2.2	Syndromes resulting from chromosomal	3L

		abnormalities	
	2.2.1	Monosomies (Turner syndrome)	
	2.2.2	Disomies and trisomies (Down Syndrome, Klinefelter's syndrome) Cri-du-chat syndrome, Philadelphia chromosome	
	2.2.3	Chromosomal Microdeletions – Prader-Willi Syndrome & Angelman Syndrome	2L
	2.3	Recognition of DNA lesions and molecular mechanism of the following DNA Repairs - Direct repair (Photoreactivation, O6 methyl guanine DNA methyl transferase)	2L
	2.3.1	Single strand repairs - Base & Nucleotide Excision Repairs, Mismatch repair (Hemimethylation of DNA)	3L
	2.3.2	Translesion synthesis and SOS repair	
	2.3.3	Recombinational repair	
	<b>3</b>	<b>Prokaryotic and eukaryotic Transcription &amp; Post-transcriptional Modifications</b>	<b>15L</b>
	3.1	Overview of Transcription	1L
	3.2.1	Role of Prokaryotic RNA polymerase and promoter; Upstream regulatory sequences	4L
	3.2.2	Stages of transcription: Initiation, elongation and termination (Rho dependent & Rho independent)	
	3.2.3	Significance of Sigma factor, Concept of Abortive initiation	
	3.3	Transcription in eukaryotes	3L
	3.3.1	Role of promoter & regulatory elements	
	3.3.2	Eukaryotic RNA polymerases and Cis acting & Trans acting elements	
	3.3.3	Mechanism of RNA transcription in eukaryotes - Formation of pre-initiation complex, initiation, elongation and termination	
	3.4	RNA Modification	1L
	r3.4.1	Mechanism of addition of 5'-cap & formation of poly A tail	
	3.4.2	Molecular mechanism of mRNA Splicing	1L
	3.4.3	RNA Processing of rRNA & tRNA	1L
	3.4.4	RNA editing - Base modifications	1L
	3.5	Role of Inhibitor -Rifampicin, Actinomycin D	1L
	3.6	Reverse transcription (Mechanism, significance & application)	2L
<b>III</b>			
<b>IV</b>	<b>4</b>	<b>Prokaryotic and eukaryotic Translation &amp; Post-translational Modifications</b>	<b>15L</b>



	4.1	Genetic basis of protein biosynthesis – Concept of structural gene & Protein ,Characteristics of Genetic code	4L
	4.2	Ribosome assembly & structure (Comparison between prokaryotic & eukaryotic ribosome)	
	4.3	tRNA – structural features and tRNA synthetase, initiator tRNA, activation of amino acids	2L
	4.4.1	Mechanism of translation in prokaryotes: Initiation, elongation & termination	3L
	4.4.2	Concept of Polyribosome	
	4.5	Mechanism of eukaryotic translation: Initiation, elongation & termination	2L
	4.6	Inhibitors of translation (prokaryotes & eukaryotes)	1L
	4.7	Post translational modifications of proteins	1L
	4.8.1	Signal hypothesis	2L
	4.8.2	Role of signal peptide & its role in Protein sorting	
	4.9	Protein localization in Nucleus	
		<b>Practicals – RPSBCHP303</b>	2 Credits
	1	Estimation of UV absorption of nucleic acids & proteins	
	2	Qualitative test for nucleic acids	
	3	Extraction of total nucleic acids from plant tissue	
	4	Study of viscosity of DNA solution	
	5	To hydrolyze DNA and separate nucleotide bases by paper chromatography	
	6	Estimation of DNA by the Diphenylamine method	
	7	Isolation of chromosomal DNA from E coli cells	
	8	Cytochemical staining of RNA by Methyl Green Pyronin	
	9	Isolation of RNA from Yeast/ Liver	
	10	Estimation of RNA by Orcinol Method	

**References:**

1. Molecular Biology of the Gene (2008) 6th ed., Watson, J.D., Baker, T.A., Bell, S.P., Gann, A., Levine, M. and Losick, R., Cold Spring Harbor Laboratory Press, Cold Spring Harbor (New York), ISBN:0-321-50781 / ISBN: 978-0-321-50781-5.
2. Cooper, G.M. and Hausman, R.E. 2009 The Cell: A Molecular Approach. 5th edition. ASM Press & Sunderland, Washington, D.C.; Sinauer Associates, MA.
3. Karp, G. 2010 Cell and Molecular Biology: Concepts and Experiments. 6 edition. John Wiley & Sons. Inc.
4. Alberts, B., Johnson,A., Lewis, J., and Enlarge, M. 2008 Molecular Biology of the Cell.
5. Genetics: Analysis & principles by Robert brooker 5th edition, McGraw-Hill Education, 201

6. Molecular Cell Biology by Lodish, Berk, Matsudaira, Kaiser, Krieger, Scott, Zipursky, Darnell

RAMNARAIN RUIA AUTONOMOUS COLLEGE

**Course Code: RPSBCH304**

**Course Title: Biology of diseases & Clinical Research****Academic year 2020-21****COURSE OUTCOMES:****After completion of the course, a student will be able to achieve these outcomes**

<b>COURSE OUTCOME</b>	<b>DESCRIPTION</b>
<b>CO 1</b>	The student will be able to understand the cooperative relationship between Biochemistry and Medicine
<b>CO 2</b>	Learn the factors in the spread of infectious diseases & Explain biology and pathogenesis of infectious agents
<b>CO 3</b>	Describe the Immune responses of body against various pathogenic organisms
<b>CO 4</b>	Recognize the Biochemical aspect of cancer, Assays for diagnosis & treatment
<b>CO 5</b>	Understand the Physiology of cardiovascular diseases
<b>CO 6</b>	Realize the importance of drug development through clinical research

## DETAILED SYLLABUS

Course Code/ Unit	Unit	Course/ Unit Title Biology of diseases & Clinical Research RPSBCH304	Credits/ Lectures 4 Credits
I	<b>1</b>	<b>Infectious diseases</b>	<b>15L</b>
	1.1	Bacterial infections: Tetanus, Diphtheria, Tuberculosis, Typhoid, Cholera	3L
	1.2	Viral infection: Measles, Mumps, influenza, HIV	
	1.3	Protozoan: Malaria and Trypanosomiasis	3L
	1.4	Parasitic infection: Leishmaniasis	
	1.5	Treatment of infectious agents	
	1.5.1	Characteristics of an ideal chemotherapeutic agent	2L
	1.6	Mode of action of antibiotics on	
	1.6.1	Cell wall (Penicillin and Cephalosporins)	1L
	1.6.2	Cell Membrane (Polymyxin and Imidazole)	1L
	1.6.3	Protein Synthesis (Streptomycin, Tetracycline and Chloramphenicol)	1L
	1.6.4	Nucleic acid (Quinolones, Nalidixic acid, Rifamycin)	1L
	1.6.5	Enzyme inhibitors (Trimethoprim)	1L
	1.7	Mechanisms of drug resistance- evolution, pathways and origin	2L
	II	<b>2</b>	<b>Cancer</b>
2.1		Genetic basis of cancer	1L
2.2		Experimental evidenced for transformation of cell	
2.3		Oncogenes	3L
2.3.1		Identification of chromosomal oncogene	
2.3.2		Gain of function mutation	
2.3.3		Conversion of proto-oncogene to oncogene	2L
2.3.4		Missense mutation, Gene amplification, chromosomal translocation, viral integration	
2.5		Tumor suppressor gene- Role of p53 and RB gene	2L
2.6		Assays – Trypan blue exclusion method, MTT assay, Soft Agar Colony Formation Assay	3L
2.7		Molecular profiling for classification of cancer,	
2.8		DNA microarray	
2.9		Cancer therapy- Antimetabolites, Chemotherapy (purine & pyrimidine analog), Demethylating agents	4L
2.9.1	Cancer immunotherapy		
III	<b>3</b>	<b>Cardiovascular diseases</b>	<b>15L</b>

	3.1	Definition; The origin of cardiovascular diseases (electrical, structural and circulatory) and types of CVDs	2L
	3.2	Defining the broad spectrum of Ailments	3L
	3.3	Stages of CVDs	
	3.4	Molecular basis of CVDs - hypertension, coronary heart (artery) disease, cerebrovascular disease, cardiomyopathy, cardiac hypertrophy, atherosclerosis, myocardial infarction.	5L
	3.5	Diagnosis and Treatment strategies: screening methods; Current treatment modalities and their advantages and disadvantages, major side effects; Challenges of treatment. biomarkers for CVDs	5L
<b>IV</b>	<b>4</b>	<b>Clinical research</b>	<b>15L</b>
	4.1	Introduction of Clinical Research	1L
	4.1.1	Clinical Trial Phases	1L
	4.1.2	Pharmacological Principal of Clinical Research Drug Development And Launch	2L
	4.1.3	Clinical Trial Design and Project Managements	2L
	4.2	Clinical Trial Design	4L
	4.2.1	Vendor Selection and Managements	
	4.2.2	Project Planning	
	4.2.3	Project Managements	
	4.2.4	Essential documents in Clinical Research and	4L
	4.3	Regulatory Requirements	
	4.3.1	Essential Documents	
	4.3.2	IND Application	
	4.3.3	Clinical Study Report	1L
	4.3.4	NDA Application	
4.3.5	Informed Consent process and Documentation		
		<b>Practicals – RPSBCHP304</b>	2 Credits
	1	WIDAL test – Qualitative & Quantitative	
	2	Antibiotic testing by agar well method	
	3	Determination of MIC of antibiotic	
	4	Separation of isoenzymes of LDH by	
	5	electrophoresis.	
	6	Visualization of cells by methylene blue	
	7	Study of viability of cells using Neutral red Assay	
	8	Case studies on clinical research	

References:

1. Textbook of Biochemistry with Clinical Correlations (2011) Devlin, T.M. John Wiley & Sons, Inc. (New York), ISBN: 978-0-4710-28173-4.
2. Immunology: A Short Course (2009) 6th ed., Coico, R and Sunshine, G., John Wiley & sons, Inc (New Jersey), ISBN: 978-0-470-08158-7
3. Biochemistry (2012) 7th ed., Berg, J.M., Tymoczko, J.L. and Stryer, L., W.H Freeman and Company (New York), ISBN: 13:978-1-4292-7635-1.
4. Genetics (2012) 6th ed., Snustad, D.P. and Simmons, M.J., John Wiley & Sons. (Singapore), ISBN: 978-1-118-09242-2.
5. Textbook of Designing Clinical Research by Stephen B. Hulley

## Modality of Assessment (SEMESTER III)

### Theory Examination Pattern:

#### A) Internal Assessment- 40%- 40 Marks

Sr No	Evaluation type	Marks
1	One Assignment/poster presentation/Model making/Quiz	20
2	One class Test (multiple choice questions / subjective)	20
	<b>TOTAL</b>	<b>40</b>

#### B) External Examination- 60%- 60 Marks

##### Semester End Theory Examination:

1. Duration - These examinations shall be of **02 ½ HOURS** duration.
2. Theory question paper pattern:

##### Paper Pattern:

Question	Options	Marks	Questions Based on
Q1. A	Any 1 out of 2	03	UNIT I
Q1. B	Any 2 out of 3	06	
Q2. A	Any 1 out of 2	03	UNIT II
Q2. B	Any 2 out of 3	06	
Q3. A	Any 1 out of 2	03	UNIT III
Q3. B	Any 2 out of 3	06	
Q4. A	Any 1 out of 2	03	UNIT IV
Q4. B	Any 2 out of 3	06	
	<b>TOTAL</b>	<b>60</b>	

### Practical Examination Pattern:

#### A) Internal Examination: 40%- 40 Marks

Particulars	Practical I, II, III & IV
Journal	05
Experimental tasks	15
<b>Total</b>	<b>20</b>

**B) External Examination: 60%- 60 Marks****Semester End Practical Examination:**

Particulars	Practical I, II, III & IV
Laboratory work	25
Viva	5
<b>Total</b>	<b>30</b>

**Overall Examination & Marks Distribution Pattern****Semester III**

Course	301			302			Grand Total
	Internal	External	Total	Internal	External	Total	
Theory	40	60	100	40	60	100	200
Practicals	20	30	50	20	30	50	100

Course	303			304			Grand Total
	Internal	External	Total	Internal	External	Total	
Theory	40	60	100	40	60	100	200
Practicals	20	30	50	20	30	50	100

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**Course Code: RPSBCH401****Course Title: Nutritional Biochemistry****Academic year 2020-21****COURSE OUTCOMES:****After completion of the course, a student will be able to achieve these outcomes**

<b>COURSE OUTCOME</b>	<b>DESCRIPTION</b>
<b>CO 1</b>	The student will learn and understand the basic concepts of nutrition, and nutritional values of foods, and Basal metabolic rate and measurement of energy requirements.
<b>CO 2</b>	Deduce the dietary requirement of carbohydrates, lipids and proteins and their biological significance
<b>CO 3</b>	The course will also aid to learn the nutritional requirement and significance of dietary minerals like macroelements and microelements
<b>CO 5</b>	Understand the condition of malnutrition, its prevention, and recommended dietary allowances.
<b>CO 6</b>	Students will be learning topics like Introduction to nutrition, Macroelements, Microelements and Nutrigenomics, Nutritional disorders and Antinutritional Factors

## DETAILED SYLLABUS

Course Code/ Unit	Unit	Course/ Unit Title Nutritional Biochemistry RPSBCH401	Credits/ Lectures 4 Credits
I	<b>1</b>	<b>Introduction to Nutrition</b>	<b>15L</b>
	1.1.1	Introduction to Human nutrition & energy supply	2L
	1.1.2	Measurement of energy content of food-	
	1.2	Calorific value of different biomolecules & mixed diet, Determination of calorific value using bomb Calorimeter (Principle & Working)	
	1.3	Respiratory quotient of food	2L
	1.4	Measurement of energy expenditure	
	1.5	Basal metabolic rate- Definition, Measurement, factors affecting BMR & its significance	
	1.5.1	Specific dynamic action of food- Definition, Mechanism & its significance	1L
	1.5.2	Sources, Daily requirement & Nutritional importance of biomolecules	2L
	1.5.3	Carbohydrates- Concept of Glycemic Index of food (Graph), Importance of fiber (Complex carbohydrate) in nutrition	
	1.5.4	Lipids-Role of essential fatty acids	
	1.6	Proteins- Essential amino acids, Nitrogen Balance (Positive, Negative Nitrogen balance & factors affecting)	4L
	1.6.1	Assessment of nutritive value of protein	
	1.6.2	Protein efficiency ratio	
	1.6.3	Biological value of protein	
	1.6.4	Net protein utilization	
	1.6.5	Chemical score	
	1.6.6	Mutual supplementation of protein	2L
	1.7	Recommended Dietary allowances (RDA)- Definition, Factors affecting RDA, RDA for adult	
1.8	Balance diet – Concept & significance, Designing diet for different subjects (infants, toddlers, adolescents, adults, geriatric, diseased state)	1L	
1.9	Numericals based on above concepts	1L	
II	<b>2</b>	<b>Macroelements</b>	<b>15L</b>
	2.1	Biochemistry of macroelements	4L
	2.2	Sources, Recommended daily allowances, Absorption, transport, excretion, Biochemical significance & Disorders related to:	

	2.2.1	Calcium	4L
	2.2.2	Phosphorous	
	2.2.3	Magnesium	3L
	2.2.4	Sodium	
	2.2.5	Potassium	
	2.2.6	Chlorine	
	2.2.7	Sulphur	4L
	2.2.8	Phosphorous	
III	<b>3</b>	<b>Microelements</b>	<b>15L</b>
	3.1	Biochemistry of microelements	
	3.2	Sources, Recommended daily allowances, Biochemical significance & Disorders related to :	4L
	3.2.1	Copper	2L
	3.2.2	Iodine	2L
	3.2.3	Manganese	1L
	3.2.4	Zinc	2L
	3.2.5	Molybdenum	1L
	3.2.6	Cobalt	1L
	3.2.7	Fluorine	1L
	3.2.8	Selenium	1L
IV	<b>4</b>	<b>Nutrigenomics, Nutritional disorders and Antinutritional Factors</b>	<b>15L</b>
	4.1	Nutrient-Gene Interaction	2L
	4.2	Drug-Nutrient Interaction	
	4.3	Obesity, Brown and White Adipose Tissue, Specific dynamic action factors affecting thermic effect of food.	1L
	4.4	Role of Leptin, Ghrenin, Adiponectin in food intake.	1L
	4.5	Pathophysiology of Nutritional disorders	1L
	4.5.1	Nutritional disorders of carbohydrate- Obesity, Non-Insulin dependent diabetes mellitus, Lactose intolerance, Celiac disease, Dental carries	2L
	4.5.2	Nutritional disorders of proteins- Protein energy Malnutrition (Kwashiorker, Marasmus & Marasmic kwashiorkor)	2L
	4.5.3	Nutritional disorders related to lipids – Essential fatty acid deficiency, cholesterol (Good vs Bad), Atherosclerosis & Arteriosclerosis	2L
	4.5.4	Eating disorders – Bulimia nervosa, Anorexia nervosa	2L
4.6	Antinutritional factors – Phytin, oxalates, tannins, trypsin inhibitors, soluble and non-soluble NSPs	2L	

**References:**

1. Textbook of Biochemistry with Clinical Correlations (2011) Devlin, T.M. John Wiley & Sons, Inc. (New York)
2. Human nutrition and dietetics by Davidson, S. et al.; Churchill Livingstone Publishers.
3. Nutrition and dietetics by Joshi, Shubhangini A.; Tata McGraw and Hill publishers
4. Nutrition Science by Srilakshmi, B.; New Age International publishers
5. Krause's Food and Nutrition Care process.(2012); Mahan, L.K Strings, S.E, Raymond, J. Elsevier's Publications.
6. The vitamins, Fundamental aspects in Nutrition and Health (2008); G.F. Coombs Jr. Elsevier's Publications..
7. Principles of Nutritional Assessment (2005) Rosalind Gibson. Oxford University Press.
8. Nutritional Biochemistry: Tom Brody.
9. Textbook of medical laboratory technology: Dr. Praful Godkar, Bhalani Publishing House
10. Biochemical methods by S Sadashivam & A Minackam, New Age International publisher.

**Course Code:** RPSBCH402

**Course Title:** Endocrinology

**Academic year 2020-21**

**COURSE OUTCOMES:**

**: After completion of the course, a student will be able to achieve these outcomes**

<b>COURSE OUTCOME</b>	<b>DESCRIPTION</b>
<b>CO 1</b>	Study the historical experiments that lead to the discovery of various hormones
<b>CO 2</b>	Deeply understand the communication between the nervous system and the endocrine system
<b>CO 3</b>	Learn the structure, functions and the disorders associated with the various hormones starting from the pituitary hormones to the gonadal hormones.
<b>CO 4</b>	Appreciate and analyze the endocrine regulation of the various metabolisms such as carbohydrate metabolism, Protein metabolism, calcium homeostasis, menstrual cycle, pregnancy and menopause.
<b>CO 5</b>	Apply the knowledge of hormones in assay of hormones such as T3, T4 and TSH and understand the strategy behind contraception.
<b>CO 6</b>	Present a case study on a hormonal and a metabolic disorder

## DETAILED SYLLABUS

Course Code/ Unit	Unit	Course/ Unit Title Endocrinology RPSBCH402	Credits/ Lectures 4 Credits
I	<b>1</b>	<b>Introduction to Endocrinology &amp; cellular signalling</b>	<b>15L</b>
	1.1	Functions of hormones and their regulation	2L
	1.2	Chemical signalling - endocrine, paracrine, autocrine, intracrine and neuroendocrine mechanisms	
	1.3	Chemical classification of hormones, transport of hormones in the circulation and their half-lives	1L
	1.4	Hormone receptors - extracellular and intracellular	3L
	1.4.1	Receptor - hormone binding, Scatchard analysis	
	1.4.2	G protein coupled receptors, G proteins, second messengers - cAMP, cGMP, IP3, DAG, Ca <sup>2+</sup>	4L
	1.5	Effector systems - adenylyl cyclase, guanylyl cyclase, PDE, PLC	
	1.5.1	Protein kinases (PKA, PKB, PKC, PKG)	
	1.5.2	Receptor tyrosine kinases - EGF, insulin and Ras - MAP kinase cascade	4L
	1.5.3	Non receptor tyrosine kinase-erythropoietin receptor JAK - STAT pathway	
	1.6	Steroid hormone Receptor	1L
	II	<b>2</b>	<b>Hypothalamic- hypophysial system and Hormones of the adrenals</b>
2.1		Hypothalamic - Pituitary axis: anatomy, histology, vasculature and secretions	2L
2.2.1		Physiological and biochemical actions of hypothalamic hormones and Anterior pituitary hormones	
2.2.2		Hormone feed- back regulatory cascade	
2.3.1		Posterior pituitary hormones –structure, physiology	3L
2.3.2		Biochemical actions of AVP and Oxytocin; Diabetes insipidus	
2.4		Histology of Adrenal Gland	2L
2.5		Physiology and action of Aldosterone; the Renin Angiotensin System	
2.6.1		Physiology and Biochemical actions of Cortisol	

	2.6.2	Regulation of cortisol synthesis: POMC and CRH	2L
	2.7.1	Adrenal medullary Hormones: Epinephrine and Norepinephrine	2L
	2.7.2	The Fight or flight response; Dual receptor hypothesis	
	2.8	General adaptation syndrome: acute and chronic stress response	3L
	2.9	Pathophysiology – Addison’s disease, Conn’s syndrome, Cushing syndrome.	
	<b>3</b>	<b>Hormones regulating Metabolism, Calcium homeostasis and Growth</b>	<b>15L</b>
III	3.1.1	Thyroid gland- Histology	3L
	3.1.2	Biosynthesis of thyroid hormone and its regulation: Role of TRH and TSH in T4 synthesis and response	
	3.1.3	Physiological and biochemical action of Thyroxine	3L
	3.1.4	Pathophysiology of thyroxine secretion: Hyper and hypothyroidism, Goitre, Graves’ disease, Cretinism, Myxoedema	
	3.2.1	Regulation of calcium homeostasis: PTH, Vitamin D and calcitonin	3L
	3.2.2	Mechanism of Ca <sup>2+</sup> regulation and pathways involving bone, skin, liver, gut and kidneys	
	3.2.3	Pathophysiology - rickets, osteomalacia, osteoporosis	3L
	3.3	Regulation of Growth: growth hormone and somatomedin	
	3.4	Endocrine disorders - gigantism, acromegaly, dwarfism, pygmies	4L
	3.5	Physiology and biochemical actions of Growth factors- EGF, PDGF and EPO	
IV	<b>4</b>	<b>Pancreatic, GI tract and Reproductive hormones</b>	<b>15L</b>
	4.1	Cells involved in release of gastrointestinal hormones	3L
	4.1.1	Gastrin family of hormones	
	4.1.2	CCK: the secretin family of hormones	
	4.1.3	Incretins; Ghrelin	
	4.2	Summary of hormone metabolite control of GI function	2L
	4.3	Hormones of the Pancreas: Structure, synthesis, physiology and biochemical actions of insulin and glucagon	

4.4	Adipocyte hormones: Adiponectin and leptin; Appetite and satiety control	3L
4.5	Pathophysiology - . Type I and type II Diabetes mellitus, Obesity and Metabolic syndrome	
4.6	Male and female sex hormones	4L
4.6.1	Interplay of hormones during ovarian and uterine phases of menstrual cycle	
4.6.2	Placental hormones; role of hormones during parturition and lactation	
4.6.3	Hormone based contraception	3L
4.6.4	Concept - ammenorrhea, menorrhagia, PMS, PCOS, Menopause	

<b>PRACTICALS</b>		
<b>RPSBCHP401 &amp; RPSBCHP402</b>	<b>Project Work</b>	4 Credits
	<p><b>GUIDELINE TO CARRY OUT PROJECTWORK</b></p> <ol style="list-style-type: none"> <li>The main purpose of introduction of Project Work at MSc II is to inculcate research culture. It will also make the students familiar with Research Methodology i.e. reference work, experimental work, data analysis of experimental data, interpretation of results obtained, writing of project work and compilation of bibliography in proper order.</li> <li>Each student shall complete a small research project during their academic year of MSc II. However, the initial reference work for the project can be started in MSc I.</li> <li><b>Nature of Research Project:</b>-Experimental-based or literature survey involving laboratory analytical work will be considered as the Research Project.</li> <li><b>Duration of Project work:</b>-Using the infrastructure available in Ramnarain Ruia Autonomous College the duration to complete the project work will be from the commencement of the project work till the end of January of MSc II (Sem IV) academic year. The duration to complete the project work from any institute apart from Ramnarain Ruia Autonomous College will be 03 months (This needs prior approval from the Department of Biochemistry, Ramnarain Ruia Autonomous College).</li> <li><b>Schedule for Submission of project Work:</b>-Experimental work or literature survey must be completed and the report on the same (2 Copies) will have to be submitted by the end of February of MSc II (Sem IV) academic year.</li> </ol>	



6. The project should be divided into the following parts:-
- Certification of completion of Project Work
  - Acknowledgement
  - Introduction
  - Review of Related Literature
  - Aims and Objectives
  - Plan of work
  - Material and Methods
  - Results
  - Discussion & Conclusion
  - Future Prospects
  - Bibliography
7. The project will be assessed.

**GUIDELINE FOR THE ASSESMENT OF PROJECT WORK**

- The practical 401 & 402 of Sem IV (Course Code No. RPSBCHP401 & RPSBCHP402) shall be exclusively devoted for the project.
- Each student will complete the project (2 copies) and get both the copies certified by the guiding teacher and the Head of Dept. (HOD) by February of MSc II (Sem IV) academic year.
- One copy of the certified project will be submitted to the Department; while the other copy will be retained by the students for his/ her personal record.
4. The candidate is required to present the Research Project to the examiner followed by Viva- Voce examination based on the project work by the examiner.
- The following Marking Scheme shall be considered while assessing the project work

Particular		Marks
a)	Project Work (Contents Submitted in the bound form)	50
b)	Presentation of Project Work to Examiner	30
c)	Viva- voce Exam based in Project Work	20
<b>TOTAL</b>		<b>100</b>

**References:**

- Lehninger: Principles of Biochemistry (2013) 6th ed., Nelson, D.L. and Cox, M.M. W.H. Freeman & Company (NewYork), ISBN:13: 978-1-4641-0962-1 / ISBN:10-14641-0962- 1.
- Vander's Human Physiology (2008) 11th ed., Widmaier, E.P., Raff, H. and Strang, K.T. McGraw Hill International Publications, ISBN: 978-0-07-128366-3.

3. Endocrinology (2007) 6th ed., Hadley, M.C. and Levine, J.E. Pearson Education (New Delhi), Inc. ISBN: 978-81-317-2610-5.
4. The Cell: A Molecular Approach (2009) 5th Ed. Cooper, G.M. and Hausman, R.E. ASM Press & Sunderland, (Washington DC), Sinauer Associates. (MA). ISBN:978-0-87893-300-6.

RAMNARAIN RUIA AUTONOMOUS COLLEGE

**Course Code:** RPSBCH403

**Course Title:** Genetic Engineering & Biotechnology

**Academic year 2020-21**

**COURSE OUTCOMES:**

After completion of the course, a student will be able to achieve these outcomes

COURSE OUTCOME	DESCRIPTION
CO 1	Acquire wide knowledge about Recombinant DNA technology by studying about various Vectors and Restriction Enzymes involved.
CO 2	Study of Various Expression Systems and Molecular Markers
CO 3	Apply the fundamental knowledge to understand Application of R-DNA technology and use of Restriction enzymes in construction of various vectors and libraries such as c-DNA & Genomic libraries
CO 4	Recognize the importance of Screening of the libraries with the help of “ Reporter Genes” and Molecular Markers such as RFLP, RAPD, AFLP.
CO 5	Analyse and Interpret the knowledge about Genetic engineering and prospects of improving crop productivity, resistance, resistance to disease and environmental stresses, methods for production of transgenic animals.
CO 6	Develop innovative methods & Apply learned knowledge to their future research

## DETAILED SYLLABUS

Course Code/ Unit	Unit	Course/ Unit Title Genetic Engineering & Biotechnology RPSBCH403	Credits/ Lectures 4 Credits	
I	<b>1</b>	<b>Introduction to RDT &amp; cloning vectors</b>	<b>15L</b>	
	1.1	Overview of RDT, Extraction and purification of plasmid and bacteriophage DNA	3L	
		Restriction and modification systems, restriction endonucleases, Concept of sticky ends, blunt ends		
	1.2	Other enzymes used in manipulating DNA molecules:	4L	
		Terminal transferases, linkers and adapters, homopolymer tailing		
	1.3	Reverse transcriptase		
	1.3.1	DNA ligase, Ligation of DNA molecules		
	1.3.2	Synthetic oligonucleotides - synthesis and use	2L	
	1.3.3	Plasmids and bacteriophages as vectors for gene cloning	4L	
	1.4	Cloning vectors based on E. coli plasmids, pBR322, pUC8, pGEM3Z		
	1.5	Cloning vectors based on M13 and $\lambda$ bacteriophage, and in vitro packaging Vectors for yeast, Ti-plasmid, and retroviral vectors, high capacity vectors		
	1.5.1	BAC and YAC	2L	
	II	<b>2</b>	<b>Gene transfer processes, selection for recombinants &amp; clone identification</b>	<b>15L</b>
		2.1	Transformation	3L
2.1.1		Preparation of competent cells using transformation and selection for transformed cells		
2.1.2		Identification for recombinants - insertional inactivation, blue-white selection Conjugation (F <sup>+</sup> & F <sup>-</sup> cells)		
2.2.1		Mechanism of conjugation in Hfr strains	3L	
2.2.2		Interrupted mating experiment technique		
2.2.3		Problems based on above concept		
2.2.4		Transduction (Generalized & Specialized)		
2.3		Introduction of phage DNA into bacterial cells	2L	
2.3.1		Identification of recombinant phages		
2.3.2		Transfection		
2.4		Transient and stable transfection	5L	
2.4.1		Chemical and physical methods of transfection		

	2.4.2	The problem of selection, direct selection, marker rescue.	
	2.4.3	cDNA and Genomic libraries, identification of a clone from gene library, colony and plaque hybridization probing, Southern and Northern hybridization	
	2.5	Methods based on detection of the translation product of the cloned gene	2L
III	<b>3</b>	<b>Expression of cloned genes, PCR &amp; DNA sequencing</b>	<b>15L</b>
	3.1.1	Vectors for expression of foreign genes in E. coli, cassettes and gene fusions	3L
	3.1.2	Challenges in producing recombinant protein in E. coli	
	3.2	Production of recombinant protein by eukaryotic cells	4L
	3.2.1	Fusion tags such as, poly-histidine, glutathione, maltose binding protein and their role in purification of recombinant proteins	
	3.3.1	Fundamentals of polymerase chain reaction	5L
	3.3.2	Types of PCR – hot start, multiplex, reverse transcriptase PCR and Nested PCR, quantitative PCR, Primer, designing for PCR, Cloning PCR products	
	3.4	DNA sequencing by Sanger's method, Automated Sanger's DNA sequencing, Pyrosequencing	3L
IV	<b>4</b>	<b>Application of genetic engineering in Biotechnology</b>	<b>15L</b>
	4.1	Site-directed mutagenesis, Protein engineering (T4-lysozyme), yeast two hybrid systems	2L
	4.2	Production of recombinant pharmaceuticals such as insulin, human growth hormone, factor VIII.	2L
	4.3	Recombinant vaccines	1L
	4.4	Gene therapy & its application	1L
	4.5	Applications in agriculture – Bt cotton, problems with genetically modified plants, glyphosate herbicide resistant crops, ethical & safety concerns	3L
	4.6	RDT in diagnosis and treatment of diseases	2L
	4.7	Model organisms: <i>Escherichia coli</i> , <i>Saccharomyces cerevisiae</i> , <i>Drosophila melanogaster</i> , <i>Caenorhabditis elegans</i> , <i>Danio rerio</i> and <i>Arabidopsis thaliana</i>	4L

	<b>Practicals – RPSBCHP403</b>	2 Credits
1	Transformation of E. coli cells with plasmid DNA.	
2	Isolation of plasmid DNA from E. coli cells.	
3	Digestion of plasmid DNA with restriction enzymes.	
4	Amplification of a DNA fragment by PCR.	
5	Complementation of $\beta$ -galactosidase for Blue and White selection.	
6	Separation of chromosomal & plasmid DNA using agarose gel electrophoresis	

**References:**

1. Gene Cloning and DNA Analysis (2010) 6th ed., Brown, T.A., Wiley-Blackwell publishing (Oxford, UK), ISBN: 978-1-4051-8173-0.
2. Principles of Gene Manipulation and Genomics (2006) 7th ed., Primrose, S.B., and Twyman, R. M., Blackwell publishing (Oxford, UK) ISBN:13: 978-1-4051-3544-3.
3. Molecular Biotechnology: Principles and Applications of Recombinant DNA (2010) 4<sup>th</sup> ed., Glick B.R., Pasternak, J.J. and Patten, C.L., ASM Press (Washington DC), ISBN: 978-1-55581-498-4 (HC).
4. Genetic engineering and its applications, P. Joshi, Botania Publishers and Distributors
5. Recombinant DNA: A short course, Watson et al, Scientific American Books
6. Biotechnology Fundamentals and Applications, S.S.Purohitt, Agrobios Publishers, 2001.

**Course Code: RPSBCH404**

**Course Title: Advanced immunology**

**Academic year 2020-21**

**COURSE OUTCOMES:**

**After completion of the course, a student will be able to achieve these outcomes**

<b>COURSE OUTCOME</b>	<b>DESCRIPTION</b>
<b>CO 1</b>	Learn the fundamental principles of immune response including molecular, biochemical and cellular basis of immune homeostasis
<b>CO 2</b>	Develop various aspects of immunological response and how its triggered and regulated
<b>CO 3</b>	Explain the specific interactions of Antigens and antibodies and the diversity of antibodies developed at the germ line DNA
<b>CO 4</b>	Complete knowledge of the molecular mechanisms and kinetics of the immune responses, both humoral and cell mediated immunity
<b>CO 5</b>	Enhance the knowledge of various immune-techniques ranging from precipitation and agglutination reactions to ELISA, Radio immunoassay and flow cytometry.
<b>CO 6</b>	The course will aid in understanding the principles of Graft rejection, Auto immunity and Antibody based therapy.

## DETAILED SYLLABUS

Course Code/ Unit	Unit	Course/ Unit Title Advanced immunology RPSBCH404	Credits/ Lectures 4 Credits
I	<b>1</b>	<b>Antigen-Antibody Interactions and Complement System</b>	<b>15L</b>
	1.1.1	Overview of Antigens and Antibodies	1L
	1.1.2	Antigen- Antibody interactions: Forces involved, antibody affinity, antibody avidity, Cross reactivity	2L
	1.1.3	Precipitation reactions – Oudins, Ouchterlony	2L
	1.1.4	Agglutination reactions: Blood typing, bacterial agglutination, passive agglutination, agglutination inhibition,	
	1.1.5	Immuno-electrophoresis: Principles of Radioimmunoassay, ELISA, Immunofluorescence, Western Blotting	3L
	1.2	Complement system	
	1.2.1	Function and components	2L
	1.2.2	Complement activation - Classical, Alternate & Lectin pathways.	3L
	1.2.3	Biological consequence of complement activation – formation of MAC	3L
II	<b>2</b>	<b>MHC, Antigen processing and presentation &amp; TCR</b>	<b>15L</b>
	2.1	Major histocompatibility complex	
	2.1.1	MHC polymorphism & organization of MHC genes- class I & class II	2L
	2.1.2	Cellular distribution & structure of class I & II molecules	
	2.1.3	MHC and immune responsiveness – Determinant-selection model and Holes-in-the-repertoire model	2L
	2.1.4	MHC and disease susceptibility (Hereditary haemochromatosis)	2L
	2.2	Antigen processing and presentation	4L
	2.2.1	Self MHC restriction of T cells	
	2.2.2	Cytosolic and endocytic pathway	
	2.2.3	Presentation of non-peptide antigens	
	2.3	T-cell Receptor	2L
	2.3.1	Structure, organization & rearrangement of TCR genes	
	2.3.2	TCR receptor complex TCR – CD3	1L



	2.3.3	TCR accessory membrane molecules	1L
	2.3.4	Ternary TCR-peptide-MHC complex	1L
III	<b>3</b>	<b>B- and T-lymphocytes</b>	<b>15L</b>
	3.1	T-cell Maturation	2L
	3.2	Thymic selection of T-cell repertoire – Positive and negative selection, central issues in thymic selection	3L
	3.3	TH-cell activation	
	3.4	T-cell differentiation	1L
	3.5	Peripheral $\gamma\delta$ T-cell	2L
	3.6	Cytotoxic T-cells	2L
	3.7	B-cell maturation	4L
	3.8	B-cell activation and proliferation – Thymus dependent and Thymus independent	
	3.9	Formation of T-B conjugates	1L
IV	<b>4</b>	<b>Cytokines &amp; Immune response in health &amp; diseases</b>	<b>15L</b>
	4.1	Humoral and cell mediated immune response	2L
	4.1.1	Cytokines - Introduction, Properties	
	4.1.2	Cytokine receptors	3L
	4.1.3	Biological functions of cytokines	
	4.1.4	Therapies based on Cytokines	
	4.2	Hypersensitivity reactions	3L
	4.2.1	Gel & Coomb's classification - types of hypersensitivity reactions	
	4.3	Transplantation immunology	3L
	4.3.1	Types of transplant; immunological basis of allograft rejection.	
	4.4	Autoimmunity	4L
	4.4.1	Organ specific –Myasthenia gravis; Hashimotos thyroiditis; Graves' Disease; Systemic – Rheumatoid arthritis, Systemic lupus erythematosus	
		<b>Practicals – RPSBCHP404</b>	2 Credits
	1	Preparation of blood smear and Differential leucocyte count.	
	2	Separation of lymphocytes by Ficoll Hypaque method	
	3	Lymphocyte viability testing by trypan blue	
	4	Assays based on precipitation reactions - Ouchterlony double immunodiffusion (DID) and Mancini radial immunodiffusion (SRID).	

	5	Assays based on agglutination reactions - Blood typing (active) & passive agglutination (C reactive protein kit & virtual lab).	
	6	Demonstration of Enzyme linked immunosorbent assay (ELISA) & DOT ELISA	
	7	Separation of serum proteins by PAGE	
	8	Virtual Lab to study immunological Techniques	

### References:

1. Kuby Immunology (2007) 6th ed., Kindt, T.L., Goldsby, R.A. and Osborne, B.A., W.H Freeman and Company (New York), ISBN:13: 978-0-7167-8590-3 / ISBN: 10:0-7617- 8590-0.
2. Immunology: A Short Course (2009) 6th ed., Coico, R and Sunshine, G., John Wiley& sons, Inc (New Jersey), ISBN: 978-0-470-08158-7.
3. Janeway's Immunobiology (2012) 8th ed., Murphy, K., Mowat, A., and Weaver, C.T., Garland Science (London & New York), ISBN: 978-0-8153-4243-4.
4. Willey, J. Sherwood L, Woolverton C, (2016), Prescott Microbiology. 10th Edition, McGraw-Hill Publisher, Columbus, OH
5. Pelczar Mi J., Chan, E.C.S., Krieg, NR, (2009). Microbiology, McGraw-Hill publisher

## Modality of Assessment (SEMESTER IV)

### Theory Examination Pattern:

#### A) Internal Assessment- 40%- 40 Marks

Sr No	Evaluation type	Marks
1	One Assignment/poster presentation/Model making/Quiz	20
2	One class Test (multiple choice questions / subjective)	20
	<b>TOTAL</b>	<b>40</b>

#### B) External Examination- 60%- 60 Marks

##### Semester End Theory Examination:

- Duration - These examinations shall be of **02 ½ HOURS** duration.
- Theory question paper pattern:

### Paper Pattern:

Question	Options	Marks	Questions Based on
Q1. A	Any 1 out of 2	03	UNIT I
Q1. B	Any 2 out of 3	06	
Q2. A	Any 1 out of 2	03	UNIT II
Q2. B	Any 2 out of 3	06	
Q3. A	Any 1 out of 2	03	UNIT III
Q3. B	Any 2 out of 3	06	
Q4. A	Any 1 out of 2	03	UNIT IV
Q4. B	Any 2 out of 3	06	
	<b>TOTAL</b>	<b>60</b>	

### Practical Examination Pattern:

#### A) Internal Examination: 40%- 40 Marks

Particulars	Practical I, II, III & IV
Journal	05
Experimental tasks	15
<b>Total</b>	<b>20</b>

**B) External Examination: 60%- 60 Marks****Semester End Practical Examination:**

Particulars	Practical I, II, III & IV
Laboratory work	25
Viva	5
<b>Total</b>	<b>30</b>

**Overall Examination & Marks Distribution Pattern****Semester IV**

Course	401			402			Grand Total
	Internal	External	Total	Internal	External	Total	
Theory	40	60	100	40	60	100	200
Practicals	20	30	50	20	30	50	100

Course	403			404			Grand Total
	Internal	External	Total	Internal	External	Total	
Theory	40	60	100	40	60	100	200
Practicals	20	30	50	20	30	50	100

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